

Folic acid (FA, pteroyl-S-glutamic acid) [75708-92-8] M 441.4, m >250°(dec), $[\alpha]_D^{25} +23^\circ$ (c 0.5, 0.1N NaOH), pK_1 2.35 (protonation N10), pK_2 2.75 (protonation N1), pK_3 3.49 (α -CO₂H), pK_4 4.65 (γ -CO₂H), pK_5 8.80 (acidic N3). If paper chromatography indicates impurities then recrystallise from hot H₂O or from dilute acid [Walker et al. *J Am Chem Soc* **70** 19 1948]. Impurities may be removed by repeated extraction with *n*-BuOH of a neutral aqueous solns of folic acid (by suspending in H₂O and adding N NaOH till the solid dissolves then adjusting the pH to ~7.0-7.5) followed by pptn with acid, filtration, and recrystn from hot H₂O. [Blakley *Biochem J* **65** 331 1975; Kalifa, Furrer, Bieri and Viscontini *Helv Chim Acta* **61** 2739 1978.] Chromatography on cellulose followed by filtration through charcoal has also been used to obtain pure acid. [Sakami and Knowles *Science* **129** 274 1959.] UV: λ_{max} 247 and 296nm (ϵ 12800 and 18700) in H₂O pH 1.0; 282 and 346nm (ϵ 27600 and 7200) in H₂O pH 7.0; 256, 284 and 366nm (ϵ 24600, 24500 and 8600) in H₂O pH 13 [Rabinowitz in *The Enzymes* (Boyer et al. Eds **2** 185 1960).

Follicle Stimulating Hormone (FSH, follitropin) [9002-68-0] M_r ~36,000. Purified by Sephadex G100 gel filtration followed by carboxymethyl-cellulose with NH₄OAc pH 5.5. The latter separates luteinising hormone from FSH. Solubility in H₂O is 0.5%. It has an isoelectric point of 4.5. A soln of 1mg in saline (100mL) can be kept at 60° for 0.5h. Activity is retained in a soln at pH 7-8 for 0.5h at 75°. The activity of a 50% aq EtOH soln is destroyed at 60° in 15 min. [Bloomfield et al. *Biochim Biophys Acta* **533** 371 1978; Hartree *Biochem J* **100** 754 1966; Pierce and Parsons *Ann Rev Biochem* **50** 465 1981.]

Fructose-1,6-diphosphate (trisodium salt) [38099-82-0] M 406.1, pK_3^{25} 6.14, pK_4^{25} 6.93 (free acid). For purification *via* the acid strychnine salt, see Neuberg, Lustig and Rothenberg [*Arch Biochem* **3** 33 1943]. The calcium salt can be partially purified by soln in ice-cold M HCl (1g per 10mL) and repptn by dropwise addition of 2M NaOH: the ppte and supernatant are heated on a boiling water bath for a short time, then filtered and the ppte is washed with hot water. The magnesium salt can be pptd from cold aqueous soln by adding four volumes of EtOH.

Fructose-6-phosphate [643-13-0] M 260.1, $[\alpha]_D^{21} +2.5$ (c 3, H₂O), pK^{25} 5.84. Crystd as the barium salt from water by adding four volumes of EtOH. The barium can be removed by passage through the H⁺ form of a cation exchange resin and the free acid collected by freeze-drying.

6-Furfurylaminopurine (Kinetin) [525-79-1] M 215.2, m 266-267°, 269-271°, 270-272°, 272° (sealed capillary), $pK_1 <1$, pK_2 3.8, pK_3 10. Platelets from EtOH and sublimes at 220°, but is best done at lower temperatures in a good vacuum. It has been extracted from neutral aqueous solns with Et₂O. [Miller et al *J Am Chem Soc* **78** 1375 1956; Bullock et al. *J Am Chem Soc* **78** 3693 1956.]

Fusaric acid (5-*n*-butylpyridine-2-carboxylic acid) [536-69-6] M 179.2, m 96-98°, 98°, 98-100°, 101-103°, pK_1 5.7, pK_2 6.16 (80% aq methoxyethanol). Dissolve in CHCl₃, dry (Na₂SO₄), filter, evaporate and recrystallise the residue from 50 parts of pet ether (b 40-60°) or EtOAc, then sublime *in vacuo*. The *copper salt* forms bluish violet crystals from H₂O and has m 258-259°. [Hardegger and Nikles *Helv Chim Acta* **39** 505 1956; Schreiber and Adam *Chem Ber* **93** 1848 1960; NMR and MS: Tschesche and Führer *Chem Ber* **111** 3500 1978.]

Fuschin (Magenta I, rosaniline HCl) [632-99-5] M 337.9, m >200°(dec). See rosaniline hydrochloride on p. 349 in Chapter 4.

D-Galactal [21193-75-9] M 146.2, m 100°, 100-102°, 104°, 103-106°, $[\alpha]_D^{20} -21.3^\circ$ (c 1, MeOH). Recryst from EtOAc, EtOH or EtOAc + MeOH. [Overend et al. *J Chem Soc* 675 1950; Wood and Fletcher *J Am Chem Soc* **79** 3234 1957; Distler and Jourdian *J Biol Chem* **248** 6772 1973.]

β -Galactosidase (from bovine testes) [9031-11-2] M_r 510,000, [EC 3.2.1.23]. Purified 600-fold by ammonium sulfate precipitation, acetone fractionation and affinity chromatography on agarose substituted with terminal thio- β -galactopyranosyl residues. [Distlern and Jourdan *J Biol Chem* 248 6772 1973.]

Gangcyclovir [9-((1,3-dihydroxy-2-propoxy)methyl)guanine; 2-amino-1,9-((2-hydroxy-1-hydroxymethyl)-ethoxymethyl)-6H-purin-6-one; Cytovene; Cymeva(e)n(e)] [82410-32-0] M 255.2, m >290°(dec), >300°(dec), monohydrate m 248-249°(dec), $pK_{Est(1)} \sim -1.1$, $pK_{Est(2)} \sim 4.1$, $pK_{Est(3)} \sim 9.7$. Recryst from MeOH. Alternatively dissolve ~90g of reagent in 700mL of distilled H₂O, filter and cool (ca 94% recovery). UV: λ_{max} in MeOH 254nm (ϵ 12,880), 270sh nm (ϵ 9040), solubility in H₂O at 25° is 4.3mg/mL at pH 7.0. **ANTIVIRAL**. [Ogilvie et al. *Can J Chem* 60 3005 1982; Ashton et al. *Biochem Biophys Res Commun* 108 1716 1982; Martin et al. *J Med Chem* 26 759 1983.]

Geranylgeranyl pyrophosphate [6699-20-3 (NH₄ salt)] M 450.5, $pK_{Est(1)} \sim <2$, $pK_{Est(2)} \sim <2$, $pK_{Est(3)} \sim 3.95$, $pK_{Est(4)} \sim 6.26$. Purified by counter-current distribution between two phases of a butanol/isopropyl ether/ammonia /water mixture (15:5:1:19) (v/v), or by chromatography on DEAE-cellulose (linear gradient of 0.02M KCl in 1mM Tris buffer, pH 8.9). Stored as a powder at 0°.

Geranyl pyrophosphate [763-10-0 (NH₄ salt)] M 314.2, $pK_{Est(1)} \sim <2$, $pK_{Est(2)} \sim <2$, $pK_{Est(3)} \sim 3.95$, $pK_{Est(4)} \sim 6.26$. Purified by paper chromatography on Whatman No 3 MM paper in a system of isopropyl alcohol/isobutyl alcohol/ammonia/water (40:20:1:39), R_F 0.77-0.82. Stored in the dark as the ammonium salt at 0°.

Gitoxigenin (3 β ,14,16 β ,21-tetrahydroxy-20(22)norcholenic acid lactone) [545-26-6] M 390.5, m 223-226°, 234°, 239-240° (anhydrous by drying at 60°), $[\alpha]_D^{20} +30^\circ$ (c 1, MeOH). Recrystn from aqueous EtOH produces plates of the sesquihydrate which dehydrate on drying at 100° *in vacuo*. It has also been recrystd from Me₂CO-MeOH and from EtOAc the crystals contain 1 mol of EtOAc with $[\alpha]_D^{21} +24.8^\circ$ (c 1, dioxane). It has UV has λ_{max} at 310, 485 and 520nm in 96% H₂SO₄. On heating with ethanolic HCl it yields *digitaligenin* with loss of H₂O. [Smith *J Chem Soc* 23 1931.]

Gliotoxin (3R-6t-hydroxy-3-hydroxymethyl-2-methyl-(5at)-2,3,6,10-tetrahydro-5aH-3,10ac-epidissulfido[1,2-a]-indol-1,4-dione) [67-99-2] M 326.4, m 191-218°(dec), 220°(dec), 221°(dec), $[\alpha]_D^{20} -254^\circ$ (c 0.6, CHCl₃), $[\alpha]_D^{25} -270^\circ$ (c 1.7, pyridine). Purified by recrystn from MeOH. Its solubility in CHCl₃ is 1%. The *dibenzoyl* derivative has m 202° (from CHCl₃-MeOH). [Glister and Williams *Nature* 153 651 1944; Elvidge and Spring *J Chem Soc Suppl* 135 1949; Johnson et al. *J Am Chem Soc* 65 2005 1943; Bracken and Raistrick *Biochem J* 41 569 1947.]

Glucose oxidase (from Aspergillus niger) [9001-37-0] M_r 186,000, [EC 1.1.3.4]. Purified by dialysis against deionized water at 6° for 48hours, and by molecular exclusion chromatography with Sephadex G-25 at room temperature. [Holt and Cotton *J Am Chem Soc* 109 1841 1987.]

Glucose-1-phosphate [59-56-3] M 260.1, $[\alpha]_D^{25} +120^\circ$ (c 3, H₂O), $[\alpha]_D^{20} +78^\circ$ (c 4, H₂O of di-K salt), pK_1 1.11, pK_2 6.13 [pK^{25} 6.50]. Two litres of 5% aq soln was brought to pH 3.5 with glacial acetic acid (+ 3g of charcoal, and filtered). An equal volume of EtOH was added, the pH was adjusted to 8.0 (glass electrode) and the soln was stored at 3° overnight. The ppt was filtered off, dissolved in 1.2L of distd water, filtered and an equal volume of EtOH was added. After standing at 0° overnight, the crystals were collected at the centrifuge, and washed with 95% EtOH, then absolute EtOH, ethanol/diethyl ether (1:1), and diethyl ether. [Sutherland and Wosilait, *J Biol Chem* 218 459 1956.] Its barium salt can be crystd from water and EtOH. Heavy metal impurities can be removed by passage of an aqueous soln (ca 1%) through an Amberlite IR-120 column (in the appropriate H⁺, Na⁺ or K⁺ forms). *Di-K salt* cryst as 2H₂O from EtOH.

Glucose-6-phosphate [acid 156-73-5; Ba salt 58823-95-3; Na salt 54010-71-8] M 260.1, m 205-207°(dec) mono Na salt, $[\alpha]_{546}^{20} +41^\circ$ (c 5, H₂O), pK_1 1.65, pK_2 6.11, pK_3^{25} 11.71 [-C₁(OH)O⁻]. Can be freed from metal impurities as described for glucose-1-phosphate. Sol of Na Salt is 5% in H₂O at 20°. Its barium salt can be purified by solution in dilute HCl and pptn by neutralising the soln. The ppt is washed with small volumes of cold water and dried in air.

Glucose-6-phosphate dehydrogenase [9001-40-5] M_r 128,000 (from Baker's yeast), 63,300 (from rat mammary gland) [EC 1.1.1.49]. The enzyme is useful for measuring pyridine nucleotides in enzyme recycling. The enzyme from Baker's yeast has been purified by $(\text{NH}_4)_2\text{SO}_4$ fractionation, Me_2CO pptn, a second $(\text{NH}_4)_2\text{SO}_4$ fractionation, concentration by DEAE-SF chromatography, a third $(\text{NH}_4)_2\text{SO}_4$ fractionation and recrystn. Crystn is induced by addition of its coenzyme NADP, which in its presence causes rapid separation of crystals at $(\text{NH}_4)_2\text{SO}_4$ concentration much below than required to ppt the amorphous enzyme. To recryst, the crystals are dissolved in 0.01M NADP (pH 7.3) with $(\text{NH}_4)_2\text{SO}_4$ at 0.55 saturation and the crystals appear within 10 to 60 min. After standing for 2-3 days (at 4°) the $(\text{NH}_4)_2\text{SO}_4$ is increased to 0.60 of saturation and more than 80% of the activity in the original crystals is recovered in the fresh crystals. [Noltmann et al. *J Biol Chem* **236** 1255 1961]. Large amounts can be obtained from rat livers. The livers are extracted with 0.025M phosphate buffer (pH 7.5), and ppted with 3M $(\text{NH}_4)_2\text{SO}_4$ (70% of activity). The ppte is dissolved in 3volumes of 0.025M phosphate (pH 7.5), dialysed against this buffer + 0.2mM EDTA at 4° for 5h, then diluted to 1% protein and the nucleic acids ppted by addition of 0.4volumes of 1% protamine sulfate. $(\text{NH}_4)_2\text{SO}_4$ is added to a concentration of 2M (pH adjusted to 7.0 with NH_3), the ppte is discarded and the supernatant is adjusted to 2.8M $(\text{NH}_4)_2\text{SO}_4$, dialysed, protein adjusted to 1% and treated with $\text{Ca}_3(\text{PO}_4)_2$ gel. The gel is added in three steps (1.5mL of 0.4% gel/mL per step) and the gel is removed by centrifugation after each addn. The third gel adsorbed 50% of the activity. The gel is eluted with 0.2M phosphate (pH 7.4, 40mL/g of gel; 60% recovery). The extract is ppted in 3volumes with $(\text{NH}_4)_2\text{SO}_4$ (adjusted to 4M) to give enzyme with an activity of 30 μ moles/mg of protein x hour. [Lowry et al. *J Biol Chem* **236** 2746 1961.] K_m values for the yeast enzyme are 20 μ M for G-6P and 2 μ M for NADP (Tris pH 8.0, 10⁻² M MgCl_2 , 38°) [Noltmann and Kuby *The Enzymes* **VII** 223 1963].

L-Glutathione (reduced form, γ -L-glutamyl-L-cysteinyl-glycine) [70-18-8] M 307.3, m 188-190°(dec), 195°(dec), $[\alpha]_D^{20}$ -20.1° (c 1, H_2O), $\text{pK}_1^{2.5}$ 2.12 (CO_2H), $\text{pK}_2^{2.5}$ 3.59 (CO_2H), $\text{pK}_3^{2.5}$ 8.75 (NH_2), $\text{pK}_4^{2.5}$ 9.65 (10.0, SH). Crystd from 50% aq EtOH, dry in a vac and store below 5°. Alternatively recrystd from aqueous EtOH under N_2 , and stored dry in a sealed container below 4°. It is soluble in H_2O . [Weygand and Geiger *Chem Ber* **90** 634 1957; Martin and Edsall *Bull Soc Chim Fr* **40** 1763 1958; *Biochem Prep* **2** 87 1952.]

L-Glutathione (oxidised) [27025-41-8] M 612.6, m 175-195°, 195°, $[\alpha]_D^{20}$ -98° (c 2, H_2O), pK_1 3.15, pK_2 4.03, pK_3 8.75. Purified by recrystn from 50% aq EtOH. Its solubility in H_2O is 5%. Store at 4°. [Li et al. *J Am Chem Soc* **76** 225 1954; Berse et al. *Can J Chem* **37** 1733 1959.]

Glutathione S-transferase (human liver) [50812-37-8] M_r 25,000, [EC 2.5.1.18]. Purified by affinity chromatography using a column prepared by coupling glutathione to epoxy-saturated Sepharose. After washing contaminating proteins the pure transferase is eluted with buffer containing reduced glutathione. The solution is then concentrated by ultrafiltration, dialysed against phosphate buffer at pH ~7 and stored in the presence of dithiothreitol (2mM) in aliquots at <-20°. [Simons and Vander Jag *Anal Biochem* **52** 334 1977.]

Glyceraldehyde-3-phosphate dehydrogenase [9001-50-7] M_r 144,000, [EC 1.2.1.12]. Purified from rabbit muscle by extraction with 0.03N KOH and ppted with $(\text{NH}_4)_2\text{SO}_4$ (0.52 of saturation). The clear supernatant was adjusted to pH 7.5 and NH_3 was added dropwise to pH 8.2-8.4. Crystals appear sometimes even without seeding. The crystals are dissolved in H_2O , filtered to remove suspended material and 2 volumes of saturated $(\text{NH}_4)_2\text{SO}_4$ at pH 8.2-8.4 is added. After 1hour the crystals appear. Recrystallise in the same way. [Cori et al. *J Biol Chem* **173** 605 1948; Furfine and Velick *J Biol Chem* **240** 844 1965, *The Enzymes* **7** 243 1963; Lui and Huskey *Biochemistry* **31** 6998 1992.] The K_m values are: NADH (3.3 μ M) and 1,3-diphosphoglycerate (8x10⁻⁷M) in pH 7.4 imidazole buffer at 26°, NAD (13 μ M), glyceraldehyde-3-P (90 μ M), P_i (2.9x10⁻⁴M), and arsenate (69 μ M) in 8.6 M NaHCO_3 buffer at 26°. [Orsi and Cleland *Biochemistry* **11** 102 1972.]

Glycerol kinase (from *Candida mycoderma*, *E coli*, rat or pigeon liver glycerokinase) [9030-66-4] M_r 251,000, [EC 2.7.1.30]. Commercial enzyme has been dialysed against 2mM Hepes, 5mM dithiothreitol and 0.3mM EDTA, followed by several changes of 20mM Hepes and 5mM dithiothreitol prior to storage under N_2 at -20°. [Knight and Cleland *Biochemistry* **28** 5728 1989.] The enzyme from pigeon liver was purified by acid-pptn (acetate buffer at pH 5.1), $(\text{NH}_4)_2\text{SO}_4$ fractionation, heat treatment (60°/ 1 h),

calcium phosphate gel filtration, a second $(\text{NH}_4)_2\text{SO}_4$ fractionation, dialysis, elution of inert proteins and crystn. This was done by repeatedly extracting the ppte from the last step with 0.05M sodium pyrophosphate (pH 7.5) containing 1mM EDTA and 0.2M $(\text{NH}_4)_2\text{SO}_4$ was added. Careful addition of solid $(\text{NH}_4)_2\text{SO}_4$ to this soln lead to crystn of the enzyme. Recrystn was repeated. The enzyme is activated by Mg^{2+} and Mn^{2+} ions and is most stable in solns in the pH 4.5-5.5 range. The stability is greatly increased in the presence of glycerol. It has K_m for glycerol of $60\mu\text{M}$ and for ATP $9\mu\text{M}$ in glycine buffer pH 9.8 and 25° . [Kennedy *Methods Enzymol* 5 476 1962.]

L-Glycerol-3-phosphate dehydrogenase (GDH, from rabbit muscle) [9075-65-4] M_r 78,000 [EC 1.1.1.8]. Recrystd by adding $(\text{NH}_4)_2\text{SO}_4$ till 0.45 saturation at pH 5.5 at 4° and the small amount of ppte is removed then satd $(\text{NH}_4)_2\text{SO}_4$ is added dropwise from time to time over several days in the cold room. The crystals are collected and recrystd until they have maximum activity. The enzyme is stable in half saturated $(\text{NH}_4)_2\text{SO}_4$ for several weeks at 4° . The equilibrium $[\text{dihydroxyacetone}][\text{NADH}][\text{H}^+]/[\text{G-3-P}][\text{NAD}]$ is $1.0 \times 10^{-12}\text{M}$ in Tris buffer at 25° . It uses NAD ten times more efficiently than NADP. The K_m for G-3-P is $1.1 \times 10^{-4}\text{M}$, for NAD it is $3.8 \times 10^{-4}\text{M}$ and for dihydroxyacetone it is $4.6 \times 10^{-4}\text{M}$ in phosphate buffer pH 7.0 and at 23.3° . Dihydroxyacetone phosphate and fructose-1,6-diphosphate are inhibitors. [Branowski *J Biol Chem* 180 515 1949, *The Enzymes* 7 85 1963; Young and Pace *Arch Biochem Biophys* 75 125 1958; Walsh and Sallach *Biochemistry* 4 1076 1965.]

L- α -Glycerol phosphocholine (Cadmium Chloride) $_x$ complex [64681-08-9] M 257.2 + (183.3) $_x$, $pK_{\text{Est}} \sim 5.5$. Glycerol phosphocholine is purified *via* the CdCl_2 complex which is purified by four recrystns from 99% EtOH by standing at 0° for 1h. The white ppte is collected, washed with EtOH, Et_2O and dried in a vacuum. The amorphous Cd complex can be converted to the crystalline form $[\text{C}_8\text{H}_{20}\text{O}_6\text{NP.CdCl}_2.3\text{H}_2\text{O}]$ by dissolving 34.4g in H_2O (410mL) and 99% EtOH (1650mL total) added slowly with stirring and allowing the clear soln to stand at 25° for 12hours, then at 5° for 12h. The crystallised complex is filtered off, washed with cold 80% EtOH and dried in air. Glycerol phosphocholine can be recovered from the complex by dissolving in H_2O (2% soln), passing through an ion-exchange column (4.9 x 100cm, of 1vol IRC-50 and 2vol of IR-45). The effluent is concentrated to a thick syrup at 45° . It is dried further at $50^\circ/\text{P}_2\text{O}_5/48\text{h}$. The vitreous product ($\sim 8.25\text{g}$) is dissolved in 99% EtOH (50mL) and the clear soln is cooled at 5° , whereby crystals appear, and then at -15° for 16h. The crystals are filtered off, washed with 99% EtOH, and Et_2O then dried at 50° in a vacuum over P_2O_5 . It can be recrystd from 99.5% EtOH, long prisms) which are *hygroscopic* and must be handled in a H_2O -free atmosphere [Tattie and McArthur *Biochem Prep* 6 16 1958; Baer and Kates *J Am Chem Soc* 70 1394 1948 ; *Acta Cryst* 21 79, 87 1966].

Glycine anhydride (2,5-diketopiperazine) [106-57-0] M 114.1, m 309-310 $^\circ$, 311-312 $^\circ$ (dec), $\sim 315^\circ$ (dec), $pK_1 -4.45$, $pK_2 -2.16$ ($pK_2 -1.94$ in AcOH). Recrystd from H_2O (plates) and can be sublimed (slowly) at 260° or at $140-170^\circ/0.5\text{mm}$. The *dihydrochloride* has m 129-130 $^\circ$, is prepd by dissolving in conc HCl and on adding EtOH to crystallisation point; dried in a vac. The *bis-1-naphthylurethane* has m 232 $^\circ$ (dec), and the *diperchlorate* has m 117 $^\circ$ (*hygroscopic*). [MS: Johnstone *J Chem Soc Perkin Trans 1* 1297 1975; NMR: Blaha and Samek *Collect Czech Chem Commun* 32 3780 1967; Sauborn *J Phys Chem* 36 179 1932; Corey *J Am Chem Soc* 60 1599 1938.]

Glycocyamine (N-guanylglycine) [352-97-6] M 117.1, m 280-284 $^\circ$ (dec), $>300^\circ$, pK^{25} 2.86 (NH_3^+). Recrystd from 15 parts of hot H_2O , or by dissolving in slightly more than the calculated amount of 2N HCl and ppting by adding an equivalent of 2N NaOH, filtering washing with cold H_2O and drying first *in vacuo* then at 60° *in vacuo*. The *hydrochloride* has m 200 $^\circ$ (dec) after recrystn from aqueous HCl as plates. The *picrate* forms needles from hot H_2O and has m 210 $^\circ$ (dec). [Brand and Brand *Org Synth Coll Vol III* 440 1955; Failey and Brand *J Biol Chem* 102 768 1933; King *J Chem Soc* 2375 1930.]

Glycodeoxycholic acid monohydrate (N-[3 α -12 α -dihydroxy-5 β -cholan-24-oyl]glycine) [360-65-6] M 467.6, m 186-177 $^\circ$ (dec), 187-188 $^\circ$, $[\alpha]_D^{23} +45.9^\circ$ (c 1, EtOH), $pK_{\text{Est}} \sim 4.4$. Recrystallises from H_2O or aqueous EtOH with 1 mol of H_2O and dried at 100° *in vacuo*. Solubility in EtOH is 5%. [UV: Lindstedt and Sjövall *Acta Chem Scand* 11 421 1957.] The *Na salt* is recrystd from EtOH/ Et_2O , m 245-250 $^\circ$, $[\alpha]_D^{23} +41.2^\circ$ (c 1, H_2O) [Wieland Hoppe Seyler's *Z Physiol Chem* 106 181 1919; Cortese *J Am Chem Soc* 59 2532 1937].

D(+)-Glycogen [9005-79-2] **M** 25,000-100,000, **m** 270-280°(dec), $[\alpha]_{546}^{20} +216^\circ$ (c 5, H₂O). A 5% aqueous soln (charcoal) was filtered and an equal volume of EtOH was added. After standing overnight at 3° the ppte was collected by centrifugation and washed with absolute EtOH, then EtOH/diethyl ether (1:1), and diethyl ether. [Sutherland and Wosilait *J Biol Chem* 218 459 1956.]

Glycogen synthase (from bovine heart) [9014-56-6] **M_r** 60,000, [EC 2.4.1.11]. Purified by pptn of the enzyme in the presence of added glycogen by polyethylene glycol, chromatography on DEAE-Sephacel and high speed centrifugation through a sucrose-containing buffer. [Dickey-Dunkirk and Kollilea *Anal Biochem* 146 199 1985.]

Gramicidin A (a pentadecapeptide from *Bacillus brevis*) [11029-61-1] **m** ~229-230°(dec). Purified by countercurrent distribution from *C₆H₆-CHCl₃, MeOH-H₂O (15:15:23:7) with 5000 tubes. Fractions were examined by UV (280nm) of small aliquots. Separation from Gramicidin C and other material occurred after 999 transfers. [Gross and Witkop *Biochemistry* 4 2495 1965; Bauer et al. *Biochemistry* 11 3266 1972.] Purified finally by recrystn from EtOH-H₂O and dried at 100°/10⁻² mm over KOH and forms platelets **m** 229-230°. Almost insoluble in H₂O (0.6%) but soluble in lower alcohols, dry Me₂CO, dioxane, acetic acid and pyridine. The commercial material is more difficult to crystallise than the synthetic compound. [Sarges and Witkop *J Am Chem Soc* 86 1861, 87 2011, 2020 1965.] It has characteristic $[\alpha]_{\text{D}}^{20} +27.3^\circ$ (c 1.3, MeOH) and UV λ_{max} 282nm (ϵ 22,100). The *N*-carbamoyldeformyl gramicidine A pptes from EtOAc-pet ether (b 40-60°).

Gramicidin C (gramicidin S, a pentadecapeptide from *Bacillus brevis*) [9062-61-7]. Same as Gramicidin A since they are isolated together and separated. [Sarges and Witkop *Biochemistry* 4 2491 1965; Hunter and Schwartz "Gramicidins" in *Antibiotics I* (Gotlieb and Shaw Eds) Springer-Verlag, NY, p.642 1967; as well as references above for Gramicidin A.]

Gramicidin S 2HCl (from *Bacillus brevis* Nagano) [15207-30-4] **M** 1214.4, **m** 277-278°(dec), $[\alpha]_{\text{D}}^{24} -289^\circ$ (c 0.4, 70% aq EtOH). Cryst in prisms from EtOH + aq HCl.

Gramicidin S [113-73-5] **M** 1141.4, **m** 268-270°, $[\alpha]_{\text{D}}^{25} -290^\circ$ (c 0.5, EtOH + 30mM aq HCl [7:3]). Crystd from EtOH. *Di-HCl* [15207-30-4] cryst from EtOH (+ few drops of HCl) has **m** 277-278°.

***N*-Guanyltiramine hydrochloride** [60-20-8] **M** 215.7, **m** 218°, **pK₁** 10.2 (phenolic OH), **pK₂** 12.4 (guanidino N). Purified on a phosphocellulose column and eluted with a gradient of aqueous NH₃ (0-10%). The second major peak has the characteristic tryptamine spectrum and is collected, lyophilised to give white crystals of the *dihydrate* which dehydrates at 100°. It has UV λ_{max} at 274.5nm (ϵ 1310) in 0.1N NaOH and 274.5nm (ϵ 1330) at pH 7.0. Excitation λ_{max} is at 280nm and emission λ_{max} is at 330nm. [Mekalanos et al. *J Biol Chem* 254 5849 1979.]

Haemoglobin A (from normal human blood) [9008-02-0] **M_r** ~64,500, **amorphous**. Purified from blood using CM-32 cellulose column chromatography. [Matsukawa et al. *J Am Chem Soc* 107 1108 1985.] For the purification of the α and β chains see Hill et al. *Biochem Prep* 10 55 1963.

Harmaline (7-methoxy-1-methyl-4,9-dihydro-3*H*- β -carboline, 4,9-dihydro-7-methoxy-1-methyl-3*H*-pyrido[3,4-*b*]indole) [304-21-2] **M** 214.3, **m** 229-230°, 229-231°, 235-237° (after distn at 120-140°/10⁻³), **pK₁** 4.2. Recrystd from MeOH and sublimed at high vacuum. It has UV in MeOH has λ_{max} 218, 260 and 376nm (log ϵ 4.27, 3.90 and 4.02 respectively); IR (Nujol) ν 1620, 1600, 1570 and 1535cm⁻¹ and in CHCl₃ ν 1470 and 1629cm⁻¹. [Spenser *Can J Chem* 37 1851 1959; Marion et al. *J Am Chem Soc* 73 305 1951; UV Prukner and Witkop *Justus Liebigs Ann Chem* 554 127 1942.] The *hydrochloride dihydrate* has **m** 234-236°(dec), the *picrate* has **m** 228-229° (sinters at 215°) from aqueous EtOH, and the *N*-acetate forms needles **m** 204-205°.

Hematin (ferrihaeme hydroxide) [15489-90-4] **M** 633.5, **m** 200°(dec), **pK_{Est}** ~ 4. Crystd from pyridine. Dried at 40° *in vacuo*.

Hematoporphyrin (3,3'-[7,12-bis-(1-hydroxyethyl)-3,8,13,17-tetramethyl-porphyrin-2,18-diyl]-dipropionic acid) [14459-29-1] M 598.7, pK_{Est} ~4.8. Purified by dissolving in EtOH and adding H₂O or Et₂O to give deep red crystals. Also recrystd from MeOH. UV has λ_{max} at 615.5, 565, 534.4 and 499.5nm in 0.1 N NaOH, and 597, 619, 634,653, 683 and 701nm in 2 N HCl. [Falk *Porphyryns and Metalloporphyryns* Elsevier, NY, p 175 1964.] It is used in the affinity chromatographic purification of Heme proteins [Olsen *Methods Enzymol* 123 324 1986.] The *O*-methyl-dimethyl ester has m 203-206° (from CHCl₃-MeOH) and the *O,O'*-dimethyl-dimethyl ester has m 145° (from CHCl₃-MeOH). [Paul *Acta Chem Scand* 5 389 1951.]

Hematoporphyrin dimethyl ester [33070-12-1] M 626.7, m 212°. Crystd from CHCl₃/MeOH.

Hematoxylin (\pm -11bc-7,11b-dihydroindeno[2,1-c]-chromen-3,4-6ar-9,10-pentaol) [517-28-2] M 302.3, m 200°(dec), 210-212°(dec). Recrystd from H₂O (as trihydrate) in white-yellow crystals which become red on exposure to light and then melt at 100-120°. It has been recrystd from Me₂CO-^{*}C₆H₆. Crystd also from dil aqueous NaHSO₃ until colourless. Soluble in alkali, borax and glycerol. Store in the dark below 0°. [Morsingh and Robinson *Tetrahedron* 26 182 1970; Dann and Hofmann *Chem Ber* 98 1498 1955.]

Hemin (ferriprotoporphyrin IX chloride) [16009-13-5] M 652.0, m sinters at 240°, pK_{Est} ~4.8. It is purified by recrystn from AcOH. Also heme (5g) is shaken in pyridine (25mL) till it dissolves, then CHCl₃ (40mL) is added, the container is stoppered and shaken for 5min (releasing the stopper occasionally). The soln is filtered under slight suction, and the flask and filter washed with a little CHCl₃ (15mL). During this period, AcOH (300mL) is heated to boiling and saturated aqueous NaCl (5mL) and conc HCl (4mL) are added. The CHCl₃ filtrate is poured in a steady stream, with stirring, into the hot AcOH mixture and set aside for 12hours. The crystals are filtered off, washed with 50% aqueous AcOH (50mL), H₂O (100mL), EtOH (25mL), Et₂O and dried in air. [Fischer *Org Synth Coll Vol III* 442 1955.]

Heparin (from pig intestinal mucosa) [9005-49-6] M_r ~3,000, amorphous, $[\alpha]_D^{20}$ ~+55° (H₂O). Most likely contaminants are mucopolysaccharides including heparin sulfate and dermatan sulfate. Purified by pptn with cetylpyridinium chloride from saturated solutions of high ionic strength. [Cifonelli and Roden *Biochem Prep* 12 12 1968.]

Heparin (sodium salt) [9041-08-1] M_r ~ 3000 (low Mol Wt, Bovine), amorphous, $[\alpha]_D^{20}$ +47° (c 1.5, H₂O). Dissolved in 0.1M NaCl (1g/100mL) and pted by addition of EtOH (150mL).

Histones (from S4A mouse lymphoma). Purification used a macroprocess column, heptafluorobutyric acid as solubilising and ion-pairing agent and an acetonitrile gradient. [McCroskey et al. *Anal Biochem* 163 427 1987.]

Hyaluronidase [9001-54-1, 37326-33-3] M_r 43,000 (bovine testes), 89,000 (bacterial), [EC 3.2.1.35]. Purified by chromatography on DEAE-cellulose prior to use. [Distler and Jourdain *J Biol Chem* 248 6772 1973.]

Hydrocortisone (11 β ,17 α ,21-trihydroxy-pregn-4-ene-3,20-dione) [50-23-7] M 362.5, m 212-213°, 214-217°, 218-221°, 220-222°, $[\alpha]_D^{22}$ +167° (c 1, EtOH). Recrystd from EtOH or isoPrOH. It is bitter tasting and has UV λ_{max} at 242 nm (log ϵ 4.20). Its solubility at 25° is: H₂O (0.28%), EtOH (1.5%), MeOH (0.62%), Me₂CO (0.93%), CHCl₃ (0.16%), propylene glycol (1.3%) and Et₂O (0.35%). It gives an intense green colour with conc H₂SO₄. [Wendler et al. *J Am Chem Soc* 72 5793 1950.]

Hydrocortisone acetate (21-acetoxy-11 β ,17 α -trihydroxy-pregn-4-ene-3,20-dione) [50-03-3] M 404.5, m 218-221.5°, 221-223°, 222-225°, $[\alpha]_D^{25}$ +166° (c 0.4, dioxane), +150.7° (c 0.5, Me₂CO). Recrystd from Me₂CO-Et₂O or aqueous Me₂CO as somewhat *hygroscopic* monoclinic crystals. UV has λ_{max} 242 nm ($A_{1cm}^{1\%}$ 390) in MeOH. Its solubility at 25° is: H₂O (0.001%), EtOH (0.45%), MeOH (0.04%), Me₂CO (1.1%), CHCl₃ (0.5%), Et₂O (0.15%) and is very soluble in Me₂NCHO. [Wendler et al. *J Am Chem Soc* 74 3630 1952; Antonucci et al. *J Org Chem* 18 7081 1953.]

(+)-Hydroquinidine anhydrous (9S-6'-methoxy-10,11-dihydrocinchonan-9-ol) [1435-55-8] **M 326.4, m 168-169°, 169°, 169-170°, 171-172°, $[\alpha]_D^{20} +231^\circ$ (c 2, EtOH), +299° (c 0.82, 0.1N H₂SO₄), $pK_{Est} \sim 8.8$** . Forms needles from EtOH and plates from Et₂O. Slightly soluble in Et₂O and H₂O but readily soluble in hot EtOH. [Heidelberger and Jacobs *J Am Chem Soc* **41** 826 1919; King *J Chem Soc* 523 1946.] The *hydrochloride* has **m 273-274°, $[\alpha]_D^{26} +184^\circ$ (c 1.3, MeOH)** and is very soluble in MeOH and CHCl₃, but less soluble in H₂O, EtOH and less soluble in dry Me₂CO. [Kyker and Lewis *J Biol Chem* **157** 707 1945; Emde *Helv Chim Acta* **15** 557 1932.]

Hydroquinine [522-66-7] **M 326.4, m 168-171°, 171.5°, $[\alpha]_D^{16} +143^\circ$ (c 1.087, EtOH), $pK^{15} 8.87$** . Recrystd from EtOH. [Rabe and Schultz *Chem Ber* **66** 120 1933.]

19-Hydroxy-4-androsten-3,17-dione [510-64-5] **M 302.4, m 167-169°, 168-170°, 169-170°, 172-173°, $[\alpha]_D^{20} +190^\circ$ (c 1, CHCl₃)**. Recrystd from Me₂CO-hexane or Et₂O-hexane. It has UV λ_{max} at 242nm in EtOH or MeOH. The *19-acetoxy* derivative has $[\alpha]_D^{26} +185^\circ$ (CHCl₃) and λ_{max} 237.5nm in EtOH. [Ehrenstein and Dünneberger *J Org Chem* **21** 774 1956.]

3-Hydroxy butyrate dehydrogenase (from *Rhodopseudomonas spheroides*) [9028-38-0] **M_r ~85,000, [EC 1.1.1.30], amorphous**. Purified by two sequential chromatography steps on two triazine dye-Sepharose matrices. [Scavan et al. *Biochem J* **203** 699 1982.]

25-Hydroxycholesterol (cholest-5-en-3 β ,25-diol) [2140-46-7] **M 402.7, m 177-179°, 178-180°, 181.5-182.5°, $[\alpha]_D^{25} -39^\circ$ (c 1.05, CHCl₃)**. Forms colourless needles from MeOH. [Schwartz *Tetrahedron Lett* **22** 4655 1981.] The *3 β -acetoxy* derivative has **m 142-142.8° (from Me₂CO), $[\alpha]_D^{25} -40.4^\circ$ (c 2, CHCl₃)**. The *3 β ,25-diacetyl* derivative has **m 119-120.5° (from MeOH), $[\alpha]_D^{25} -35.5^\circ$ (CHCl₃)**. [Dauben and Bradlow *J Am Chem Soc* **72** 4248 1950; Ryer et al. *J Am Chem Soc* **72** 4247 1950.]

18-Hydroxy-11-deoxycorticosterone (18,21-dihydroxypregn-4-en-3,20-dione tautomeric with 18,20-epoxy-20,21-dihydroxypregn-4-en-3-one) [379-68-0] **M 346.5, m 168-170°, 171-173°, 191-195°, 200-205°, $[\alpha]_D^{20} +151^\circ$ (c 1, CHCl₃)**. Recrystn from Et₂O-Me₂CO gave crystals **m 200-205°**, when recrystd from Me₂CO it had **m 191-195°**. It has UV λ_{max} at 240nm. The *21-O-acetoxy-18-hydroxy* derivative has **m 158-159° (from Et₂O-^{*}C₆H₆)** and the *21-O-acetoxy-18,20-epoxy* derivative has **m 149-154° (from Et₂O)**. [Kahnt et al. *Helv Chim Acta* **38** 1237 1955; Pappo *J Am Chem Soc* **81** 1010 1959.]

R-(-)-2-Hydroxy-3,3-dimethyl- γ -butyrolactone (3-hydroxy-4,4-dimethyl-4,5-dihydrofuran-2-one, D-pantolactone) [599-04-2] **M 130.1, m 89-91°, 90.5-91.5°, 91°, 92-93°, b 120-122°/15mm, $[\alpha]_D^{20} -28^\circ$ (c 5, MeOH), $[\alpha]_D^{20} -51^\circ$ (c 3, H₂O)**. Recrystallise from Et₂O-pet ether, diisopropyl ether or ^{*}C₆H₆-pet ether and sublime at 25°/0.0001mm. It hydrolyses readily to the hydroxy-acid and racemises when heated above 145°. The *Brucine salt* has **m 211-212° (from EtOH)**. [Kuhn and Wieland *Chem Ber* **73** 1134 1940; and Stiller et al. *J Am Chem Soc* **62** 1779 1940; Bental and Tishler *J Am Chem Soc* **68** 1463 1946.]

(±)-Ibotenic acid monohydrate (α -[3-hydroxy-5-isoxazolyl]-glycine, α -amino-3-hydroxy-5-isoxazoleacetic acid) [2552-55-8] **M 176.1, m 144-146° (monohydrate), 151-152° (anhydrous), 148-151°, pK_1 2, pK_2 5.1, pK_3 8.2**. It has been converted to the ammonium salt (**m 121-123° dec**) dissolved in H₂O and passed through an Amberlite IR 120 resin (H⁺ form) and eluted with H₂O. The acidic fractions were collected, evaporated to dryness and the residue recrystd from H₂O as the monohydrate (**m 144-146°**). The anhydrous acid is obtained by making a slurry with MeOH, decanting and evaporating to dryness and repeating the process twice more to give the anhydrous acid (**m 151-152°**). Recrystn from H₂O gives the monohydrate. [Nakamura *Chem Pharm Bull Jpn* **19** 46 1971.] The *ethyl ester* forms needles when crystd from a small volume of Et₂O and has **m 78-79° and IR (CHCl₃) with ν 3500-2300 (OH), 1742 (ester CO), 1628, 1528cm⁻¹, and UV with λ_{max} (EtOH) at 206nm (ϵ 7080)**. The *hydrazide* has **m 174-175° (from MeOH) with IR (KBr) 1656 (C=O)cm⁻¹**.

2-Iminothiolane hydrochloride (2-iminotetrahydrothiophene) [4781-83-3] **M 137.6, m 187-192°**, 190-195°, 193-194°, 202-203°, **pK <2 (free base)**. Recryst from MeOH-Et₂O (**m** 187-192°) but after sublimation at ~180°/0.2mm the melting point rose to 202-203°. It has NMR with δ 2.27 (2H, t), 3.25 (2H, t) and 3.52 (2H, t) in (CD₃)₂SO. [King et al. *Biochemistry* 17 1499 1978.] The *free base* is purified by vacuum distn (**b** 71-72°/6mm) with IR (film) with ν 1700 (C=N)cm⁻¹ and NMR (CDCl₃) with δ at 3.58 (2H, t) and 2.10-2.8 (4H, m). The *free base* is stable on storage but slowly hydrolyses in aqueous solns with half lives at 25° of 390h at pH 9.1, 210h at pH 10 and 18 h at pH 11. [Alagon and King *Biochemistry* 19 4343 1980.]

trans-Indol-3-ylacrylic acid [1204-06-4] **M 187.2, m 190-195°(dec), 195°(dec), 196°(dec), 195-196°(dec), pK_{Est} ~ 4.2**. Recrystd from AcOH, H₂O or EtOAc-cyclohexane. UV in MeOH has λ_{\max} at 225, 274 and 325nm. [Shaw et al. *J Org Chem* 23 1171 1958; constitution: Rappe *Acta Chem Scand* 18 818 1964; Moffatt *J Chem Soc* 1442 1957; Kimming et al. *Hoppe Seyler's Z Physiol Chem* 371 234 1958.]

3-Indolylbutyric acid [133-32-4] **M 203.2, m 120-123°, 123-125°, 124°, pK 4.84**. Recrystd from H₂O. It is soluble in EtOH, Et₂O and Me₂CO but insoluble in CHCl₃. [Bowman and Islip *Chem Ind London* 154 1971; Jackson and Manske *J Am Chem Soc* 52 5029 1930; Albaum and Kaiser *Am J Bot* 24 420 1937.] UV has λ_{\max} 278 and 320nm in isoPrOH [Elvidge *Quart J Pharm Pharmacol* 13 219 1940]. The *methyl ester* has **m** 73-74° (from *C₆H₆-pet ether) and **b** 230°/6mm [Bullock and Hand *J Am Chem Soc* 78 5854 1951]. Also recrystd from EtOH/water [James and Ware *J Phys Chem* 89 5450 1985].

3-Indolylpyruvic acid [392-12-1] **M 203.2, m~210°(dec), 208-210°(dec), 219°(dec), pK_{Est} ~ 2.4**. Recrystd from Me₂CO-*C₆H₆, EtOAc-CHCl₃, Me₂CO-AcOH (crystals with 1 molecule of AcOH) and dioxane-*C₆H₆ (with 0.5 molecule of dioxane) [Shaw et al. *J Org Chem* 23 1171 1958; Kaper and Veldstra *Biochim Biophys Acta* 30 401 1958]. The *ethyl ester* has **m** 133° (from Et₂O) and its *2,4-dinitrophenylhydrazone* has **m** 255° (from Me₂CO). [Baker *J Chem Soc* 461 1946.]

myo-Inositol (cyclohexane[1r,2c,3c,4t,5c,6t]-hexol) [87-89-8] **M 180.2, m 218° (dihydrate), 225-227°, 226-230°**. Recrystd from aq 50% ethanol or H₂O forming a dihydrate, or anhydrous crystals from AcOH. The dihydrate is efflorescent and becomes anhydrous when heated at 100°. The anhydrous crystals are not hygroscopic. Solubility in H₂O at 25° is 14%, at 60° it is 28%, slightly soluble in EtOH but insoluble in Et₂O. [Ballou and Anderson *J Am Chem Soc* 75 748 1953; Anderson and Wallis *J Am Chem Soc* 70 2931 1948.]

Interferons [α IFN, β IFN and γ IFN]. Interferons are a family of glycosylated proteins and are cytokines which are produced a few hours after cells have been infected with a virus. Interferons protect cells from viral infections and have antiviral activities at very low concentrations (~3 x 10⁻⁴ M, less than 50 molecules are apparently sufficient to protect a single cell). Double stranded RNA are very efficient inducers of IFNs. There are three main types of IFNs. The α IFNs are synthesised in lymphocytes and the β IFNs are formed in infected fibroblasts. The α and β families are fairly similar consisting of ca 166 to 169 amino acids. Although γ IFNs are also small glycosylated proteins (ca 146 amino acids), they are different because they are not synthesised after viral infections but are produced by lymphocytes when stimulated by **mitogens** (agents that induced cell division).

Several of these IFNs of mouse and human lymphocytes and fibroblasts are available commercially and have been best prepared in quantity by recombinant DNA procedures because they are produced in very small amounts by the cells. The commercial materials do not generally require further purification for their intended purposes. [Pestkas, Interferons and Interferon standards and general abbreviations, *Methods Enzymol*, Wiley & Sons, 119 1986, ISBN 012182019X; Lengyel, Biochemistry of interferons and their actions, *Ann Rev Biochem* 51 251-282 1982; De Maeyer and De Maeyer-Guignard, Interferons in *The Cytokine Handbook*, 3rd Edn, Thomson et al. Eds, pp. 491-516 1998 Academic Press, San Diego, ISBN 0126896623.]

Interleukin (from human source). Purified using lyophilisation and desalting on a Bio-Rad P-6DC desalting gel, then two steps of HPLC, first with hydroxylapatite, followed by a TSK-125 size exclusion column. [Kock and Luger *J Chromatogr* 296 293 1984.]

Interleukin-2 (recombinant human) [94218-72-1] $M_r \sim 15,000$, amorphous. Purified by reverse phase HPLC. [Weir and Sparks *Biochem J* **245** 85 1987; Robb et al. *Proc Natl Acad Sci USA* **81** 6486 1984.]

Interleukins (IL-1, IL-2 —IL18). Interleukins are cytokines which cause a variety of effects including stimulation of cell growth and proliferation of specific cells, e.g. stem cells, mast cells, activated T cells, colony stimulating factors etc, as well as stimulating other ILs, prostaglandins release etc. They are small glycosylated proteins (ca 15 kD, 130-180 amino acids produced from longer precursors) and are sometimes referred to by other abbreviations, e.g. IL-2 as TCGF (T cell growth factor), IL-3 as multi-CSF (multilineage colony stimulating factor, also as BPA, HCSEF, MCSF and PSF). They are produced in very small amounts and are commercially made by recombinant DNA techniques in bacteria or Sf21 insect cells. Interleukins for human (h-IL), mouse (m-IL) and rat (r-IL) are available and up to IL-18 are available commercially in such purity that they can be used directly without further refinement, particularly those that have been obtained by recombinant DNA procedures which are specific. As well as the interleukins, a variety of antibodies for specific IL reactions are available for research or IL identification. [Symons et al. *Lymphokines and Interferons, A Practical Approach*, Clemens et al. Eds, p. 272 1987; IRL Press, Oxford, ISBN 1852210354, 1852210362; Thomson et al. Eds, *The Cytokine Handbook*, 3rd Edn, 1998; Academic Press, San Diego, ISBN 0126896623.]

Iodonitrotetrazolium chloride (2[4-iodophenyl]-3-[4-nitrophenyl]-5-phenyl-2H-tetrazolium chloride) [146-68-9] M 505.7, m 229°(dec), ~245°(dec). Recrystd. from H₂O, aqueous EtOH or EtOH-Et₂O. Alternatively dissolve in the minimum volume of EtOH and add Et₂O; or dissolve in hot H₂O (charcoal), filter and ppt by adding conc HCl. Filter solid off and dry at 100°. Solubility in H₂O at 25° is 0.5%, and in hot MeOH-H₂O (1:1) it is 5%. [Fox and Atkinson *J Am Chem Soc* **72** 3629 1950.]

Iodonitrotetrazolium violet-Formazan [7781-49-9] M 471.3, m 185-186°. Dissolve in boiling dioxane (20g in 300mL), add H₂O (100mL) slowly, cool, filter and dry *in vacuo* at 100°. Its solubility in CHCl₃ is ~1%. [UV: Fox and Atkinson *J Am Chem Soc* **72** 3629 1950.]

5-Iodouridine (5-iodo-1-[β-D-ribofuranosyl]-pyrimidine-2,4(1H)-dione) [1024-99-3] M 370.1, m 205-208°(dec), 210-215°(dec), $[\alpha]_D^{20}$ -23.5° (c 1, H₂O), pK^{20} 8.5. Recrystd from H₂O and dried *in vacuo* at 100°. UV has λ_{max} 289nm (0.01N HCl) and 278nm (0.01N NaOH). [Prusoff et al. *Cancer Res* **13** 221 1953.]

3-Isobutyl-1-methylxanthine (3-isobutyl-1-methylpurine-2,6-dione) [28822-58-4] M 222.3, m 199-210°, 202-203°, $pK_{Est} \sim 6.7$ (acidic NH). Recrystd from aqueous EtOH.

Isopentenyl pyrophosphate [358-71-4] M 366.2, $pK_{Est(1)} \sim <2$, $pK_{Est(2)} \sim <2$, $pK_{Est(3)} \sim 3.95$, $pK_{Est(4)} \sim 6.26$. Purified by chromatography on Whatman No 1 paper using *tert*-butyl alcohol/formic acid/water (20:5:8, R_F 0.60) or 1-propanol/ammonia/water (6:3:1. R_F 0.48). Also purified by chromatography on a DEAE-cellulose column or a Dowex-1 (formate form) ion-exchanger using formic acid and ammonium formate as eluents. A further purification step is to convert it to the monocyclohexylammonium salt by passage through a column of Dowex-50 (cyclohexylammonium form) ion-exchange resin. Can also be converted into its lithium salt.

DL-Isoserine (±-3-amino-2-hydroxypropionic acid) [632-12-2] M 105.1, m 250-252°(dec), 235°(dec), 237°(dec), 245°(dec), pK_1^{25} 2.78 (acidic), pK_2^{25} 9.27 (basic). Recrystd from H₂O or 50% aqueous EtOH. It has an isoelectric pH of 6.02. [Rinderknocht and Niemann *J Am Chem Soc* **75** 6322 1953; Gundermann and Holtmann *Chem Ber* **91** 160 1958; Emerson et al. *J Biol Chem* **92** 451 1931.] The *hydrobromide* has m 128-130° (from aqueous HBr) [Schöberl and Braun *Justus Liebigs Ann Chem* **542** 288 1939].

Isoxanthopterin (2-amino-4,7-dihydroxypteridine) [529-69-1] M 179.4, $m > 300^\circ$, pK_1^{20} -0.5 (basic), pK_2^{20} 7.34 (acidic), pK_3^{20} 10.06 (acidic). Purified by repeated pptn from alkaline solutions by acid (preferably AcOH), filter, wash well with H₂O then EtOH and dried at 100°. Purity is checked by paper chromatography [R_F 0.15 (*n*-BuOH, AcOH, H₂O, 4:1:1); 0.33 (3% aq NH₄OH)]. [Goto et al. *Arch Biochem*

Biophys 111 8 1965.] For biochemistry see Blakley *Biochemistry of Folic Acid and Related Pteridines* North Holland Publ Co, Amsterdam 1969.]

Kanamycin B (Bekanamycin, 4-*O*-[2,6-diamino-2,6-dideoxy- α -D-glucopyranosyl]-6-*O*-[3-amino-3-deoxy- α -D-glucopyranosyl]-2-deoxystreptomine) [4696-76-8, 29701-07-3 (sulfate salt)] **M** 483.5, **m** 170-179^o(dec), 178-182^o(dec), $[\alpha]_D^{18} +130^{\circ}$ (c 0.5, H₂O), **pK** 7.2. A small quantity (24mg) can be purified on a small Dowex 1 x 2 column (6 x 50mm), the correct fraction is evapd to dryness and the residue crystd from EtOH containing a small amount of H₂O. [Umezawa et al. *Bull Chem Soc Jpn* 42 537 1969.] It has been crystd from H₂O by dissolving ~1g in H₂O (3mL), adding Me₂NCHO (3mL) setting aside at 4^o overnight, The needles are collected and dried to constant weight at 130^o. It has also been recrystd from aq EtOH. It is slightly sol in CHCl₃ and isoPrOH. [IR: Wakazawa et al. *J Antibiot* 14A 180, 187 1961; Ito et al. *J Antibiot* 17 A 189 1964.]

Lactate dehydrogenase (from dogfish, Beef muscle) [9001-60-9] **M_r** 140,000 [EC 1.1.1.27]. 40-Fold purification by affinity chromatography using Sepharose 4B coupled to 8-(6-aminoethyl)amino-5'-AMP or -NAD⁺. [Lees et al. *Arch Biochem Biophys* 163 561 1974; Pesce et al. *J Biol Chem* 239 1753 1964.]

Lactoferrin (from human whey). Purified by direct adsorption on cellulose phosphate by batch extraction, then eluted by a stepped salt and pH gradient. [Foley and Bates *Anal Biochem* 162 296 1987.]

Lecithin (1,2-diacylphosphatidylcholine mixture) [8002-43-5] **M** ~600-800, **amorphous**. From hen egg white. Purified by solvent extraction and chromatography on alumina. Suspended in distilled water and kept frozen until used [Lee and Hunt *J Am Chem Soc* 106 7411 1984, Singleton et al. *J Am Oil Chem Soc* 42 53 1965]. For purification of commercial egg lecithin see Pangborn [*J Biol Chem* 188 471 1951].

Lectins (proteins and/or glycoproteins of non-immune origin that agglutinate cells, from seeds of *Robinia pseudoacacia*), **M** ~100,000. Purified by pptn with ammonium sulfate and dialysis; then chromatographed on DE-52 DEAE-cellulose anion-exchanger, hydroxylapatite and Sephacryl S-200. [Wantyghem et al. *Biochem J* 237 483 1986.]

Leucopterin (2-amino-5,8-dihydropteridine-4,6,7(1*H*)-trione) [492-11-5] **M** 195.1, **m** >300^o (dec), **pK₁²⁰** -1.66, **pK₂²⁰** 7.56, **pK₃²⁰** 9.78, **pK₄²⁰** 13.6. Purified by dissolving in aqueous NaOH, stirring with charcoal, filtering and precipitating by adding aqueous HCl, then drying at 100^o in a vacuum. It separates with 0.5 moles of H₂O. Its solubility in H₂O is 1g/750 litres [Albert et al. *J Chem Soc* 4219 1952; Albert and Wood *J Appl Chem (London)* 2 591 1952; Pfleiderer *Chem Ber* 90 2631 1957].

DL- α -Lipoamide (\pm -6,8-thioctic acid amide, 5-[1,2]-dithiolan-3-ylvaleric acid amide) [3206-73-3] **M** 205.3, **m** 124-126^o, 126-129^o, 130-131^o. Recrystd from EtOH and has UV with λ_{\max} 331nm in MeOH. [Reed et al. *J Biol Chem* 232 143 1958; IR: Wagner et al. *J Am Chem Soc* 78 5079 1956.]

DL- α -Lipoic acid (\pm -6,8-thioctic acid, 5-[1,2]-dithiolan-3-ylvaleric acid) [1077-28-7] **M** 206.3, **m** 59-61^o, 60.5-61.5^o and 62-63^o, **b** 90^o/10⁻⁴mm, 150^o/0.1mm, **pK²⁵** 4.7. It forms yellow needles from cyclohexane or hexane and has been distd at high vacuum, and sublimes at ~90^o and very high vacuum. Insoluble in H₂O but dissolves in alkaline soln. [Lewis and Raphael *J Chem Soc* 4263 1962; Soper et al. *J Am Chem Soc* 76 4109; Reed and Niu *J Am Chem Soc* 77 416 1955; Tsuji et al. *J Org Chem* 43 3606 1978; Calvin *Fed Proc USA* 13 703 1954.] The *S*-benzylthiouronium salt has **m** 153-154^o (evacuated capillary; from MeOH), 132-134^o, 135-137^o (from EtOH). The *d*- and *l*- forms have **m** 45-47.5^o and $[\alpha]_D^{23} \pm 113^{\circ}$ (c 1.88, *C₆H₆) and have UV in MeOH with λ_{\max} at 330nm (ϵ 140).

Lipoprotein lipase (from bovine skimmed milk) [9004-02-8] [EC 3.1.1.34]. Purified by affinity chromatography on heparin-Sepharose [Shirai et al. *Biochim Biophys Acta* **665** 504 1981].

Lipoproteins (from human plasma). Individual human plasma lipid peaks were removed from plasma by ultracentrifugation, then separated and purified by agarose-column chromatography. Fractions were characterised immunologically, chemically, electrophoretically and by electron microscopy. [Rudel et al. *Biochem J* **13** 89 1974.]

Lipoteichoic acids (from gram-positive bacteria) [56411-57-5]. Extracted by hot phenol/water from disrupted cells. Nucleic acids that were also extracted were removed by treatment with nucleases. Nucleic resistant acids, proteins, polysaccharides and teichoic acids were separated from lipoteichoic acids by anion-exchange chromatography on DEAE-Sephacel or by hydrophobic interaction on octyl-Sepharose [Fischer et al. *Eur J Biochem* **133** 523 1983].

D-Luciferin (firefly luciferin, S-2[6-hydroxybenzothiazol-2-yl]-4,5-dihydrothiazol-4-carboxylic acid), [2591-17-5] **M 280.3**, **m 189.5-190°(dec)**, **196°(dec)**, **201-204°**, **205-210°(dec, browning at 170°)**, $[\alpha]_D^{22} -36^\circ$ (c 1.2, DMF), $pK_{Est(1)} \sim 1.2$ (benzothiazole-N), $pK_{Est(2)} \sim 1.6$ (thiazolidine-N), $pK_{Est(3)} \sim 6.0$ (CO₂H), $pK_{Est(4)} 8.5$ (6OH). Recrystallises as pale yellow needles from H₂O, or MeOH (83mg from 7mL). It has UV λ_{max} at 263 and 327nm (log ϵ 3.88 and 4.27) in 95% EtOH. The Na salt has a solubility of 4mg in 1 mL of 0.05M glycine. [White et al. *J Am Chem Soc* **83** 2402 11961, **85** 337 1963; UV and IR: Bitler and McElroy *Arch Biochem* **72** 358 1957; Review: Cormier et al. *Fortschr Chem Org Naturst* **30** 1 1973.]

Lumiflavin (7,8,10-trimethylbenzo[g]pteridine-2,4(3H,10H)-dione) [1088-56-8] **M 256.3**, **m 330°(dec)**, **340°(dec)**, **pK 10.2**. Forms orange crystals upon recrystn from 12% aqueous AcOH, or from formic acid. It sublimes at high vacuum. It is freely soluble in CHCl₃, but not very soluble in H₂O and most organic solvents. In H₂O and CHCl₃ soln it has a green fluorescence. UV has λ_{max} at 269, 355 and 445nm (ϵ 38,800, 11,700 and 11,800 respectively) in 0.1N NaOH and 264, 373 and 440nm (ϵ 34,700, 11,400 and 10,400 respectively) in 0.1N HCl while UV in CHCl₃ has λ_{max} at 270, 312, 341, 360, 420, 445 and 470nm. [Hemmerich et al. *Helv Chim Acta* **39** 1242 1956; Holiday and Stern *Chem Ber* **67** 1352 1834; Yoneda et al. *Chem Pharm Bull Jpn* **20** 1832 1972; Birch and Moye *J Chem Soc* 2622 1958; Fluorescence: Kuhn and Moruzzi *Chem Ber* **67** 888 1934.]

Magnesium protoporphyrin dimethyl ester [14724-63-1] **M 580.7**. Crude product dissolved in as little hot dry *C₆H₆ as possible and left overnight at room temperature to crystallise. [Fuhrhop and Graniek *Biochem Prep* **13** 55 1971.]

α -Melanotropin [581-05-5] (**13 amino acids peptide**), $[\alpha]_D^{25} -58.5^\circ$ (c 0.4, 10% aq AcOH). Extract separated by ion-exchange on carboxymethyl cellulose, desalted, evapd and lyophilised, then chromatographed on Sephadex G-25. [Lande et al. *Biochem Prep* **13** 45 1971.]

β -Melanotropin. [9034-42-8] (**18-22 amino acids peptide**), **amorphous**. Extract separated by ion-exchange on carboxymethyl cellulose, desalted, evapd and lyophilised, then chromatographed on Sephadex G-25. [Lande et al. *Biochem Prep* **13** 45 1971.]

6-Mercaptopurine monohydrate [6112-76-1] **M 170.2**, **m 314-315°(dec)**, **~315°(dec)**, **313-315°(dec)**, $pK_1^{20} 7.77$, $pK_2^{20} 10.84$. Recrystallises from H₂O as yellow crystals of the monohydrate which become anhydrous on drying at 140°. It has UV λ_{max} at 230 and 312nm (ϵ 14,000 and 19,600) in 0.1N NaOH; 222 and 327nm (ϵ 9,2400 and 21,300), and 216 and 329nm (ϵ 8,740 and 19,300) in MeOH. [Albert and Brown *J Chem Soc* 2060 1954; IR: Brown and Mason *J Chem Soc* 682 1957; UV: Fox et al. *J Am Chem Soc* **80** 1669 1958; UV: Mason *J Chem Soc* 2071 1954.]

6-Mercaptopurine-9- β -D-ribofuranoside [574-25-4] M 284.3, m 208-210°(dec), 210-211°(dec), 220-223°(dec), 222-224°(dec), $[\alpha]_D^{25}$ -73° (c 1, 0.1N NaOH), pK 7.56. Recrystd from H₂O or EtOH. It has UV λ_{\max} in H₂O at 322nm (pH 1), 320 nm (pH 6.7) and 310nm (pH 13). [IR: Johnson et al. *J Am Chem Soc* 80 699 1958; UV: Fox et al. *J Am Chem Soc* 80 1669 1958.]

Metallothionein (from rabbit liver) [9038-94-2]. Purified by precipitation to give Zn- and Cd-containing protein fractions and running on a Sephadex G-75 column, then isoelectric focusing to give two protein peaks [Nordberg et al. *Biochem J* 126 491 1972].

Methadone hydrochloride (2-dimethylamino-4-ethoxycarbonyl-4,4-diphenylbutane HCl) [1095-90-5] M 345.9, m 241-242°, pK₁²⁵ 8.94, pK₂²⁰ 10.12 (free base). Crystd from EtOH.

Methoxantin coenzyme (PQQ, pyrrolo quinoline quinone, 2,7,9-tricarboxy-1H-pyrrolo-[2,3-f]-quinoline-4,5-dione, 4,5-dihydro-4,5-dioxo-1H-pyrrolo[2,3-f]quinoline-2,7,9-tricarboxylic acid) [72909-34-3] M 330.2, m 220°(dec). Efflorescent yellow-orange needles on recrystn from H₂O by addition of Me₂CO, or better from a supersaturated aqueous soln, as it forms an acetone adduct. [Forrest et al. *Nature* 280 843 1979.] It has also been purified by passage through a C-18 reverse phase silica cartridge or a silanised silica gel column in aqueous soln whereby methoxantin remains behind as a red-orange band at the origin. This band is collected and washed thoroughly with dilute aqueous HCl (pH 2) and is then eluted with MeOH-H₂O (7:3) and evapd *in vacuo* to give the coenzyme as a red solid. It has also been purified by dissolving in aqueous 0.5M K₂CO₃ and acidified to pH 2.5 whereby PQQ pptes as a deep red solid which is collected and dried *in vacuo*. Methoxantin elutes at 3.55 retention volumes from a C18 μ Bondapak column using H₂O-MeOH (95:5) + 0.1% AcOH pH 4.5. It has UV λ_{\max} at 247 and 330nm (shoulder at 270nm) in H₂O and λ_{\max} at 250 and 340nm in H₂O at pH 2.5. With excitation at λ_{ex} 365nm it has a λ_{\max} emission at 483nm. The ¹³C NMR has δ : 113.86, 122.76, 125.97, 127.71, 130.68, 137.60, 144.63, 146.41, 147.62, 161.25, 165.48, 166.45, 173.30 and 180.00.

When a soln in 10% aqueous MeCO is adjusted to pH 9 with aqueous NH₃ and kept at 25° for 30 min, the *acetone adduct* is formed; UV has λ_{\max} at 250, 317 and 360nm (H₂O, pH 5.5) and with λ_{ex} at 360nm it has max fluorescence at λ_{\max} at 465nm; and the ¹³C NMR [(CD₃)₂SO, TMS] has δ : 29.77, 51.06, 74.82, 111.96, 120.75, 121.13, 125.59, 126.88, 135.21, 139.19, 144.92, 161.01, 161.47, 165.17, 168.61, 190.16 and 207.03. It also forms a *methanol adduct*.

When it is reacted with Me₂SO₄-K₂CO₃ in dry Me₂NCHO at 80° for 4h, it forms the *trimethyl ester* which has m 265-267°(dec) [260-263°(dec)] after recrystn from hot MeCN (orange crystals) with UV λ_{\max} at 252 and 344nm (H₂O) and 251, 321 and 373nm (in MeOH; MeOH adduct ?). [Duine et al. *Eur J Biochem* 108 187 1980; Duine et al. *Adv Enzymology* 59 169 1987; Corey and Tramontano *J Am Chem Soc* 103 5599 1981; Gainor and Weinreb *J Org Chem* 46 4319 1981; Hendrickson and de Vries *J Org Chem* 17 1148 1982; McKenzie, Moody and Reese *J Chem Soc Chem Commun* 1372 1983.]

Methyl benzylpenicillinate [653-89-4] M 348.3, m 97°, $[\alpha]_D^{20}$ +328° (c 1, MeOH). Crystd from CCl₄.

5-Methylphenazinium methyl sulfate [299-11-6] M 306.3, m 155-157° (198°dec by rapid heating). It forms yellow prisms from EtOH (charcoal). Solubility in H₂O at 20° is 10%. In the presence of aqueous KI it forms a *semiquinone* which crystallises as blue leaflets from EtOH. [Wieland and Roseen *Chem Ber* 48 1117 1913; Voriskova *Collect Czech Chem Commun* 12 607 1947; Bülow *Chem Ber* 57 1431 1924.]

1-Methyl-4-phenyl-1,2,3,6-tetrahydropyridine hydrochloride (MPTP) [23007-85-4] M 209.7, m 196-198°, pK_{Est} ~ 9.3. Purified by recrystn from Me₂CO + isoPrOH. The *free base* has b 137-142°/0.8 mm, n_D^{25} 1.5347. [Schmidle and Mansfield *J Am Chem Soc* 78 425 1956; Defeudis *Drug Dev Res* 15 1 1988.]

6- α -Methylprednisolone (Medrol, 11 β ,17-21-trihydroxy-6 α -methylpregna-1,4-dien-3,20-dione) [83-43-2] M 347.5, m 226-237°, 228-237°, 240-242°, $[\alpha]_D^{24}$ +91° (c 0.5, dioxane). Recrystd from EtOAc. UV has λ_{\max} in 95% EtOH 243nm (ϵ 14,875). The *21-acetoxy derivative* has m 205-

208° (from EtOAc), $[\alpha]_D^{24} +95^\circ$ (c 1, CHCl₃). [Spero et al. *J Am Chem Soc* **78** 6213 1956; Fried et al. *J Am Chem Soc* **81** 1235 1959; ¹H NMR: Slomp and McGarvey *J Am Chem Soc* **81** 2200 1959.]

5-Methyltetrahydrofolic acid disodium salt (prefolic A) [68792-52-9] M 503.4, pK₁ 2.4 (N10 protonation), pK₂ 2.7 (pyrimidine N1 protonation), pK₃ 3.5 (α-CO₂H), pK₄ 4.9 (γ-CO₂H), pK₅ 5.6 (N5-Me), pK₆ 8.5 (3NHCO acidic). Check purity by measuring UV at pH 7.0 (use phosphate buffer) and it should have λ_{\max} 290nm and λ_{\min} 245nm with a ratio of A₂₉₀/A₂₅₀ of 3.7. This ratio goes down to 1.3 as oxidation to the dihydro derivative occurs. The latter can be reduced back to the tetrahydro compound by reaction with 2-mercaptoethanol at room temp. If oxidation had occurred then the compound should be chromatographed on DEAE-cellulose (~0.9 milliequiv/g, in AcO⁻ form) in (NH₄)₂CO₃ (1.5 M) and washed with 1M NH₄OAc containing 0.01M mercaptoethanol till free from UV absorption and then washed with 0.01M mercaptoethanol. All is done in a nitrogen atmosphere. The reduced folate is then eluted with a gradient between 0.01M mercaptoethanol and 1M NH₄OAc containing 0.01M mercaptoethanol and the fractions with absorption at 290nm are collected. These are evapd under reduced pressure at 25° and traces of NH₄OAc and H₂O are removed at high vacuum/25° (~24-48h). The residue is dissolved in the minimum volume of 0.01M mercaptoethanol and an equivalent of NaOH is added to convert the acid to the diNa salt and evaporated to dryness at high vacuum/25°. The product should have λ_{\max} 290nm (ε 32,000) in pH 7.0 buffer. [Sakami *Biochem Prep* **10** 103 1963.]

5-Methyltryptamine hydrochloride (3-[2-aminoethyl]-5-methylindole hydrochloride) [1010-95-3] M 210.7, m 289-291°(dec), 290-292°, pK_{Est(1)} ~ -3 (protonation of ring NH), pK_{Est(2)} ~ 9.0 (CH₂NH₂), pK_{Est(3)} ~ 10.9 (acidic indole NH). Recrystd from H₂O. The *free base* has m 93-95° (from *C₆H₆-cyclohexane), and the *picrate* has m 243°(dec) (from EtOH). [Young *J Chem Soc* 3493 1958; Gaddum et al. *Quart J Exp Physiol* **40** 49 1955; Röhm *Hoppe Seyler's Z Physiol Chem* **297** 229 1954.]

4-Methylumbelliferone(β) hydrate (7-hydroxy-4-methylcoumarin) [90-33-5] M 194.2, m 185-186°, 185-188°, 194-195°, pK_{Est} ~ 10.0 (phenolic OH). Purified by recrystn from EtOH. It is insoluble in cold H₂O, slightly soluble in Et₂O and CHCl₃, but soluble in MeOH and AcOH. It has blue fluorescence in aqueous EtOH, and has UV λ_{\max} 221, 251 and 322.5nm in MeOH. IR has ν 3077 br, 1667, 1592, 1385, 1267, 1156, 1130 and 1066 cm⁻¹. The *acetate* has m 153-154°. [Woods and Sapp *J Org Chem* **27** 3703 1962.]

4-Methylumbellifer-7-yl-α-D-glucopyranoside [17833-43-1] M 338.3, m 221-222°, $[\alpha]_D^{20} 237^\circ$ (c 3, H₂O). Recrystd from hot H₂O.

4-Methylumbellifer-7-yl-β-D-glucopyranoside [18997-57-4] M 338.3, m 210-212°, 211°, $[\alpha]_D^{20} -61.5^\circ$ (c 2, pyridine), -89.5° (c 0.5, H₂O for half hydrate). Recrystallises as the half hydrate from hot H₂O. [Constantzas and Kocourek *Collect Czech Chem Commun* **24** 1099 1959; De Re et al. *Ann Chim (Rome)* **49** 2089 1959.]

1-Methyluric acid [708-79-2] M 182.1, m >350°, pK₁ 5.75 (basic), pK₂ 10.6 (acidic). Recrystd from H₂O. [Bergmann and Dikstein *J Am Chem Soc* **77** 691 1955.] It has UV λ_{\max} at 231 and 283. nm (pH 3) and 217.5 and 292.5nm (pH >12) [Johnson *Biochem J* **5** 133 1952].

Mevalonic acid lactone [674-26-0] M 130.2, m 28°, b 145-150°/5mm. Purified *via* the *dibenzylethylenediammonium salt* (m 124-125°) [Hofmann et al. *J Am Chem Soc* **79** 2316 1957], or by chromatography on paper or on Dowex-1 (formate) column. [Bloch et al. *J Biol Chem* **234** 2595 1959.] Stored as *N,N'*-dibenzylethylenediamine (DBED) salt, or as the lactone in a sealed container at 0°.

Mevalonic acid 5-phosphate [1189-94-2] M 228.1, pK_{Est(1)} ~ 1.5 (PO₄H₂), pK_{Est(2)} ~ 4.4 (CO₂H), pK_{Est(3)} ~ 6.31 (PO₄H⁻). Purified by conversion to the *tricyclohexylammonium salt* (m 154-156°) by treatment with cyclohexylamine. Crystd from water/acetone at -15°. Alternatively, the phosphate was chromatographed by ion-exchange or paper (Whatman No 1) in a system isobutyric acid/ammonia/water (66:3:30; R_F 0.42). Stored as the cyclohexylammonium salt.

Mevalonic acid 5-pyrophosphate [1492-08-6] M 258.1, $pK_{Est(1)} \sim <2$, $pK_{Est(2)} \sim <2$, $pK_{Est(3)} \sim 3.95$ (PO_4), $pK_{Est(4)} 4.4$ (CO_2H), $pK_{Est(5)} \sim 6.26$ (PO_4). Purified by ion-exchange chromatography on Dowex-1 formate [Bloch et al. *J Biol Chem* **234** 2595 1959], DEAE-cellulose [Skilletar and Kekwick, *Anal Biochem* **20** 171 1967], on by paper chromatography [Rogers et al. *Biochem J* **99** 381 1966]. Likely impurities are ATP and mevalonic acid phosphate. Stored as a dry powder or as a slightly alkaline (pH 7-9) soln at -20° .

Mithramycin A (Aureolic acid, Plicamycin) [18378-89-7] M 1085.2, m 180-183°, $[\alpha]_D^{20} -51^\circ$ (c 0.3, EtOH), $pK_{Est} \sim 9.2$. Purified from $CHCl_3$, and is soluble in MeOH, EtOH, Me_2CO , EtOAc, Me_2SO and H_2O , and moderately soluble in $CHCl_3$, but is slightly soluble in *C_6H_6 and Et_2O . Fluorescent antitumour agent used in flow cytometry. [Thiem and Meyer *Tetrahedron* **37** 551 1981; NMR: Yu et al. *Nature* **218** 193 1968.]

Mitomycin C [50-07-7] M 334.4, m $>360^\circ$, $pK_{Est(2)} \sim 8.0$. Blue-violet crystals form *C_6H_6 -pet ether. It is soluble in Me_2CO , MeOH and H_2O , moderately soluble in *C_6H_6 , CCl_4 and Et_2O but insoluble in pet ether. It has UV λ_{max} at 216, 360 and a weak peak at 560nm in MeOH. [Stevens et al. *J Med Chem* **8** 1 1965; Shirahata and Hirayama *J Am Chem Soc* **105** 7199 1983.]

Muramic acid [R-2(2-amino-2-deoxy-D-glucose-3-yloxy)-propionic acid] [1114-41-6] M 251.2, m 145-150°(dec), 152-154°(dec), 155°(dec), $[\alpha]_D^{25} +109^\circ$ (c 2, H_2O), $+165.0^\circ$ (extrapolated to 0 time) $\rightarrow +123^\circ$ [after 3h (c 3, H_2O)], $pK_{Est(1)} \sim 3.8$ (CO_2), $pK_{Est(2)} \sim 7.7$ (NH_2). It has been recrystd from H_2O or aqueous EtOH as monohydrate which loses H_2O at 80° *in vacuo* over P_2O_5 . Sometimes contains some NaCl. It has been purified by dissolving 3.2g in MeOH (75mL), filtered from some insoluble material, concentrated to ~ 10 mL and refrigerated. The colourless crystals are washed with absolute MeOH. This process does not remove NaCl; to do so the product is recrystd from a equal weight of H_2O to give a low yield of very pure acid (0.12g). On paper chromatography 0.26 μ g give one ninhydrin positive spot after development with 75% phenol (R_F 0.51) or with *sec*-BuOH- HCO_2O - H_2O (7:1:2) (R_F 0.30). [Matsushima and Park *Biochem Prep* **10** 109 1963; *J Org Chem* **27** 3581 1962.] The acid has been also purified by dissolving 990mg in 50% aqueous EtOH (2mL), cooling, collecting the colourless needles on a sintered glass funnel and dried over P_2O_5 at $80^\circ/0.1$ mm to give the anhydrous acid. [Lambert and Zilliken *Chem Ber* **93** 2915 1960.] Alternatively the acid is dissolved in a small volume of H_2O , neutralised to pH 7 with ion exchange resin beads (IR4B in OH^- form), filtered, evaporated and dried. The residue is recrystd from 90% EtOH (v/v) and dried as above for 24h. [Strange and Kent *Biochem J* **71** 333 1959.] The *N*-acetyl derivative has m $\sim 125^\circ$ (dec) and $[\alpha]_D^{20} +41.2^\circ$ after 24h (c 1.5, H_2O). [Watanabe and Saito *J Bacteriol* **144** 428 1980.]

Muscimol (pantherine, 5-aminoethyl-3[2h]-isoxazolone) [2763-96-4] M 114.1, m 170-172°(dec), 172-174°(dec), 172-175°, 175°, 176-178°(dec), $pK_{Est(1)} \sim 6$ (acidic, ring 2-NH), $pK_{Est(2)} \sim 8$ ($CH_2CH_2NH_2$). Recrystd from MeOH-tetrahydrofuran or EtOH and sublimed at 110-140° (bath) at 10^{-4} mm and gives a yellow spot with ninhydrin which slowly turns purple [NMR: Bowden et al. *J Chem Soc (C)* **172** 1968]. Also purified by dissolving in the minimum volume of hot H_2O and adding EtOH dropwise until cloudy, cool, and colourless crystals separate; IR: ν 3445w, 3000-2560w br, 2156w, 1635s and 1475s cm^{-1} . [NMR: Jager and Frey *Justus Liebigs Ann Chem* **817** 1982.] Alternatively it has been purified by two successive chromatographic treatments on Dowex 1 x 8 with the first elution with 2M AcOH and a second with a linear gradient between 0—2M AcOH and evaporating the desired fractions and recrystallising the residue from MeOH. [McCarry and Savard *Tetrahedron Lett* **22** 5153 1981; Nakamura *Chem Pharm Bull Jpn* **19** 46 1971.]

Mycophenolic acid (6-[1,3-dihydro-7-hydroxy-5-methoxy-4-methyl-1-oxoisobenzofuran-6-yl]-4-methylhex-4-enoic acid) [24280-93-1] M 320.3, m 141°, 141-143°, $pK_{Est(1)} \sim 2.5$ (CO_2H), $pK_{Est(2)} \sim 9.5$ (phenolic OH). Purified by dissolving in the minimum volume of EtOAc, applying to a silica gel column (0.05-0.2 mesh) and eluting with a mixture of EtOAc + $CHCl_3$ + AcOH (45:55:1) followed by recrystn from heptane-EtOAc, from aqueous EtOH or from hot H_2O and drying *in vacuo*. It is a weak dibasic acid moderately soluble in Et_2O , $CHCl_3$ and hot H_2O but weakly soluble in *C_6H_6 and

toluene. [Birch and Wright *Aust J Chem* **22** 2635 1969; Canonica et al. *J Chem Soc Perkin Trans 1* 2639 1972; Birkinshaw, Raistrick and Ross *Biochem J* **50** 630 1952.]

Myoglobin (from sperm whale muscle). [9047-17-0] M_r ~17,000. Purified by CM-cellulose chromatography and Sephadex G-50 followed by chromatography on Amberlite IRC-50 Type III or BioRex 70 (<400mesh). The crystalline product as a paste in saturated $(\text{NH}_4)_2\text{SO}_4$ at pH 6.5-7.0 may be stored at 4° for at least 4 years unchanged, but must not be kept in a freezer. [Anres and Atassi *Biochemistry* **12** 942 1980; Edmundson *Biochem Prep* **12** 41 1968.]

Myricetin (Cannabiscetin, 3,3',4',5,5',7-hexahydroxyflavone) [529-44-2] M 318.2, m >300°, 357°(dec) (polyphenolic $pK_{\text{Est}} \sim 8-11$). Recrystd from aq EtOH (m 357° dec, as monohydrate) or Me_2CO (m 350° dec, with one mol of Me_2CO) as yellow crystals. Almost insol in CHCl_3 and AcOH. The hexaacetate has m 213°. [Hergert *J Org Chem* **21** 534 1956; Spada and Camerani *Gazzetta* **86** 965, 975 1956; Kalff and Robinson *J Chem Soc* **127** 181 1925.]

Nalidixic acid (1-ethyl-7-methyl-1,8-naphthyridin-4-one-3-carboxylic acid) [389-08-2] M 232.3, m 226.8-230.2°, 228-230°, 229-230°, pK 6.0. Crystd from H_2O or EtOH as a pale buff powder. It is soluble at 23° in CHCl_3 (3.5%), toluene (0.16%), MeOH (0.13%), EtOH (0.09%), H_2O (0.01% and Et_2O (0.01%). It inhibits nucleic acid and protein synthesis in yeast. [Leshner et al. *J Med and Pharm Chem* **5** 1063 1962.]

Naloxone hydrochloride hydrate (Narcan, 1-N-propenyl-7,8-dihydro-14-hydroxymorphinan-6-one hydrochloride) [51481-60-8] M 399.9, m 200-205°, $[\alpha]_D^{20}$ -164° (c 2.5, H_2O), $pK_{\text{Est}(1)} \sim 6$ (N-propenyl), $pK_{\text{Est}(2)} \sim 9.6$ (phenolic OH). This opiate antagonist has been recrystd from EtOH + Et_2O or H_2O . It is soluble in H_2O (5%) and EtOH but insoluble in Et_2O . The free base has m 184° (177-178°) after recrystn from EtOAc, $[\alpha]_D^{20}$ -194.5° (c 0.93, CHCl_3). [Olofson et al. *Tetrahedron Lett* 1567 1977; Gold et al. *Med Res Rev* **2** 211 1982.]

Naltrexone hydrochloride dihydrate (1-N-cyclopropylmethyl-7,8-dihydro-14-hydroxymorphinan-6-one hydrochloride) [16676-29-2] M 413.9, m 274-276°, $[\alpha]_D^{20}$ -173° (c 1, H_2O), $pK_{\text{Est}(1)} \sim 6$ (N-cyclopropylmethyl), $pK_{\text{Est}(2)} \sim 9.6$ (phenolic OH). This narcotic antagonist has been purified by recrystn from MeOH and dried air. The free base has m 168-170° after recrystn from Me_2CO . [Cone et al. *J Pharm Sci* **64** 618 1975; Gold et al. *Med Res Rev* **2** 211 1982.]

α -Naphthoflavone (7,8-benzoflavone) [604-59-1] M 272.3, m 153-155°, 155°, pK 8-9 (phenolic OH). Recrystd from EtOH or aqueous EtOH. [IR: Cramer and Windel *Chem Ber* **89** 354 1956; UV Pillon and Massicot *Bull Soc Chim Fr* 26 1954; Smith *J Chem Soc* 542 1946; Mahal and Venkataraman *J Chem Soc* 1767 1934.] It is a competitive inhibitor of human estrogen synthase. [Kellis and Vickery *Science* **225** 1032 1984.]

Naphthol AS-acetate (3-acetoxynaphthoic acid anilide) [1163-67-3] M 305.3, m 152°, 160°. Recrystd from hot MeOH and dried *in vacuo* over P_2O_5 . It is slightly soluble in AcOH, EtOH, CHCl_3 or $^*\text{C}_6\text{H}_6$. It is a fluorogenic substrate for albumin esterase activity. [Chen and Scott *Anal Lett* **17** 857 1984.] At λ_{ex} 320nm it had fluorescence at λ_{em} 500nm. [Brass and Sommer *Chem Ber* **61** 1000 1928.]

1-Naphthyl phosphate disodium salt [2183-17-7] M 268.1, pK_1^{26} 0.97, pK_2^{26} 5.85 (for free acid). Purified through an acid ion-exchange column (in H^+ form) to give the free acid which is obtained by freeze drying and recrystn from Me_2CO + $^*\text{C}_6\text{H}_6$, or by adding 2.5 vols of hot CHCl_3 to a hot soln of 1 part acid and 1.2 parts Me_2CO and cooling (m 155-157°, 157-158°). The acid is dissolved in the minimum volume of H_2O to which 2 equivalents of NaOH are added and then freeze dried, or by adding the equivalent amount of MeONa in MeOH to a soln of the acid in MeOH and collecting the Na salt, washing with cold MeOH then Et_2O and drying in a vacuum. [Friedman and Seligman *J Am Chem Soc* **72** 624 1950; Chanley and Feageson *J*

Am Chem Soc 77 4002 1955.] It is a substrate for alkaline phosphatase [Gomori *Methods Enzymol* 4 381 1957, 128 212 1968], and prostatic phosphatase [Babson *Clin Chem* 30 1418 1984]. See entry on p. 444.

2-Naphthyl phosphate monosodium salt [14463-68-4] M 246.2, m 296° (sintering at 228°), pK_1^{26} 1.28, pK_2^{26} 5.53 (for free acid). The free acid is purified as for the preceding 1-isomer and has m 176-177°, 177-178° after recrystn from $CHCl_3 + Me_2CO$ as the 1-isomer above. It is neutralised with one equivalent of NaOH and freeze dried or prepared as the 1-isomer above. Its solubility in H_2O is 5%. It also forms a 0.5 Na.1 H_2O salt which has m 203-205° (244° ?). [Friedman and Seligman *J Am Chem Soc* 72 624 1950; Chanley and Fegeason *J Am Chem Soc* 77 4002 1955.] See entry on p. 444 in Chapter 5.

D(+)-Neopterin [2009-64-5] M 253.2, m >300°(dec), $[\alpha]_{546}^{20} +64.5^\circ$ (c 0.14, 0.1M HCl), $[\alpha]_D^{25} +50.1^\circ$ (c 0.3, 0.1N HCl), pK_1 2.23 (basic), pK_2 7.89 (acidic). Purified as biopterin. Also purified on a Dowex 1 x 8 (formate form) column and eluted with 0.03M ammonium formate buffer pH 8.0 then pH 7.2. The fluorescent neopterin fraction is evapd under reduced pressure leaving neopterin and ammonium formate (the latter can be sublimed out at high vacuum). The residue is stirred for 24h with EtOH and the solid is collected and recrystd from H_2O [Viscontini et al. *Helv Chim Acta* 53 1202 1970; see Wachter et al. Eds *Neopterin* W de Guyter, Berlin 1992].

β -Nicotinamide adenine dinucleotide (diphosphopyridine nucleotide, NAD, DPN) [53-84-9] M 663.4, $[\alpha]_D^{23} -34.8^\circ$ (c 1, H_2O), pK_1 2.2 (PO_4H), pK_2 4.0 (adenine NH_2), pK_3 6.1 (PO_4^-). Purified by paper chromatography or better on a Dowex-1 ion-exchange resin. The column was prepared by washing with 3M HCl until free of material absorbing at 260nm, then with water, 2M sodium formate until free of chloride ions and, finally, with water. NAD, as a 0.2% soln in water, adjusted with NaOH to pH 8, was adsorbed on the column, washed with water, and eluted with 0.1M formic acid. Fractions with strong absorption at 360nm were combined, acidified to pH 2.0 with 2M HCl, and cold acetone (ca 5L/g of NAD) was added slowly and with constant agitation. It was left overnight in the cold, then the ppte was collected in a centrifuge, washed with pure acetone and dried under vacuum over $CaCl_2$ and paraffin wax shavings [Kornberg *Methods Enzymol* 3 876 1957]. Purified by anion-exchange chromatography [Dalziel and Dickinson *Biochemical Preparations* 11 84 1966.] The purity is checked by reduction to NADH (with EtOH and yeast alcohol dehydrogenase) which has ϵ_{340nm} 6220 $M^{-1}cm^{-1}$. [Todd et al. *J Chem Soc* 3727,3733 1957.] [pK_a , Lamborg et al. *J Biol Chem* 231 685 1958.] The free acid crystallises from aq Me_2CO with 3 H_2O and has m 140-142°. It is stable in cold neutral aqueous solns in a desiccator ($CaCl_2$) at 25°, but decomposes at strong acid and alkaline pH. Its purity is checked by reduction with yeast alcohol dehydrogenase and EtOH to NADH and noting the OD at 340nm. Pure NADH has ϵ_{340} 6.2 x 10⁴ $M^{-1}cm^{-1}$, i.e. 0.1 μ mole of NADH in 3mL and in a 1cm path length cell has an OD at 340nm of 0.207.

β -Nicotinamide adenine dinucleotide reduced di-Na salt trihydrate (reduced diphosphopyridine nucleotide sodium salt, NADH) [606-68-8] M 763.5, pK as for NAD. This coenzyme is available in high purity and it is advised to buy a fresh preparation rather than to purify an old sample as purification will invariably lead to a more impure sample contaminated with the oxidised form (NAD). It has UV λ_{max} at 340nm (ϵ 6,200 $M^{-1}cm^{-1}$) at which wavelength the oxidised form NAD has no absorption. At 340 nm a 0.161mM solution in a 1cm (pathlength) cell has an absorbance of 1.0 unit. The purity is best checked by the ratio $A_{280nm}/A_{340nm} \sim 2.1$, a value which increases as oxidation proceeds. The dry powder is stable indefinitely at -20°. Solutions in aqueous buffers at pH ~7 are stable for extended periods at -20° and for at least 8h at 0°, but are oxidised more rapidly at 4° in a cold room (e.g. almost completely oxidised overnight at 4°). [UV: Drabkin *J Biol Chem* 175 563 1945; Fluorescence: Boyer and Thorell *Acta Chem Scand* 10 447 1956; Redox: Rodkey *J Biol Chem* 234 188 1959; Schlenk in *The Enzymes* 2 250, 268 1951; Kaplan in *The Enzymes* 3 105, 112 1960.] Deuterated NADH, i.e. NADD, has been purified through the anion exchange resin AG-1 x 8 (100-200 mesh, formate form) and through a Bio-Gel P-2 column. [Viola, Cook and Cleland *Anal Biochem* 96 334 1979.]

β -Nicotinamide adenine dinucleotide phosphate (NADP, TPN) [53-59-8] M 743.4, pK_1 1.1 (PO_4H_2), pK_2 4.0 (adenine NH_2), pK_3 6.1 (PO_4^-). Purified by anion-exchange chromatography in much the same way as for NAD [Dalziel and Dickinson *Biochem J* 95 311 1965; *Biochemical Preparations* 11 87 1966]. Finally it is purified by dissolving in H_2O and precipitating with 4 volumes of Me_2CO and dried in

vacuo over P₂O₅. It is unchanged by storing *in vacuo* at 2°. [Hughes et al. *J Chem Soc* 3733 1957, Schuster and Kaplan *J Biol Chem* 215 183 1955.] Deuterated NADPH, i.e. NADPD, has been purified through the anion exchange resin AG-1 x 8 (100-200 mesh, formate form) and through a Bio-Gel P-2 column. λ_{\min} 259nm (ϵ 18.000) at pH 7.0. [Viola, Cook and Cleland *Anal Biochem* 96 334 1979.]

β -Nicotinamide adenine dinucleotide phosphate reduced tetrasodium salt (reduced diphosphopyridine nucleotide phosphate sodium salt, NADPH) [2646-71-1] M 833.4, pK as for NADP. Mostly similar to NADH above.

β -Nicotinamide mononucleotide (NMN) [1094-61-7] M 334.2, $[\alpha]_{\text{D}}^{23}$ -38.3° (c 1, H₂O), pK_{Est} ~ 6.1 (PO₄⁻). Purified by passage through a Dowex 1 (Cl⁻ form), washed with H₂O until no absorbance at 260 nm. The tubes containing NMN are pooled, adjusted to pH 5.5-6 and evapd *in vacuo* to a small volume. This is adjusted to pH 3 with dilute HNO₃ in an ice bath and treated with 20 volumes of Me₂CO at 0-5°. The heavy white ppte is collected by centrifugation at 0°. It is best stored wet and frozen or can be dried to give a gummy residue. It has λ_{\max} 266nm (ϵ 4600) and λ_{\min} 249nm (ϵ 3600) at pH 7.0 (i.e. no absorption at 340nm). It can be estimated by reaction with CN⁻ or hydrosulfite which form the 4-adducts equivalent to NADH) which has UV λ_{\max} 340nm (ϵ 6200). Thus after reaction an OD₃₄₀ of one is obtained from a 0.1612mM soln in a 1cm path cuvette. [Plaut and Plaut *Biochem Prep* 5 56 1957; Maplan and Stolzenbach *Methods Enzymol* 3 899 1957; Kaplan et al. *J Am Chem Soc* 77 815 1955.]

(-)-Nicotine (1-methyl-2[3-pyridyl]-pyrrolidine) [54-11-5] M 162.2, b 123-125°/17mm, 246.1°/730.5mm, 243-248°/atm (partial dec), d_4^{20} 1.097, n_{D}^{20} 1.5280, $[\alpha]_{\text{D}}^{20}$ -169° (c 1, Me₂CO), pK₁¹⁵ 6.16 (pyridine N⁺), pK₂¹⁵ 10.96 (pyrrolidine N⁺). Very pale yellow *hygroscopic* oil with a characteristic odour (tobacco extract) with browns in air on exposure to light. Purified by fractional distn under reduced pressure in an inert atmosphere. A freshly distd sample should be stored in dark sealed containers under N₂. It is a strong base, at 0.05 M soln it has a pH of 10.2. Very soluble in organic solvents. It is soluble in H₂O and readily forms salts. [UV: Parvis *J Chem Soc* 97 1035 1910; Dobbie and Fox *J Chem Soc* 103 1194 1913.] The *hydrochlorides* (mono- and di-) form deliquescent crystals soluble in H₂O and EtOH but insoluble in Et₂O. It has also been purified *via* the ZnCl₂ double salt. [Ratz *Monatsh Chem* 26 1241 1905; Biosynthesis: Nakan and Hitchinson *J Org Chem* 43 3922 1978.] The *picrate* has m 218° (from EtOH). **POISONOUS.**

(±)-Nicotine [22083-74-5] M 162.2, b 242.3°/atm, d_4^{20} 1.082 (pK see above). Purified by distn. Its solubility in EtOH is 5%. The *picrate* forms yellow needles from hot H₂O and has m 218°. The *methiodide* has m 219° (from MeOH).

Nisin [1414-45-5] M 3354.2. Polypeptide from *S. lactis*. Crystd from EtOH. [Berridge et al. *Biochem J* 52 529 1952; synthesis by Fukase et al. *Tetrahedron Lett* 29 795 1988.]

2-Nitrophenyl- β -D-galactopyranoside [369-07-3] M 301.3, m 185-190°, 193°, 193-194°, $[\alpha]_{\text{D}}^{18}$ -51.9° (c 1, H₂O). Purified by recrystn from EtOH. [Seidman and Link *J Am Chem Soc* 72 4324 1950; Snyder and Link *J Am Chem Soc* 75 1758 1953]. It is a chromogenic substrate for β -galactosidases [Jagota et al. *J Food Sci* 46 161 1981].

4-Nitrophenyl- α -D-galactopyranoside [7493-95-0] M 301.3, m 166-169°, 173°, $[\alpha]_{\text{D}}^{25}$ +248 (c 1, H₂O). Purified by recrystn from H₂O or aqueous EtOH. The *monohydrate* has m 85° which resolidifies and melts at 151-152° (the hemihydrate) which resolidifies and melts again at 173° as the anhydrous form. Drying the monohydrate at 60° yields the hemihydrate and drying at 100° gives the anhydrous compound. The *tetraacetate* has m 147° after drying at 100°. [Jermyn *Aust J Chem* 15 569 1962; Helfreich and Jung *Justus Liebigs Ann Chem* 589 77 1954.] It is a substrate for α -galactosidase [Dangelmaier and Holmsen *Anal Biochem* 104 182 1980].

4-Nitrophenyl- β -D-galactopyranoside [3150-24-1] M 301.3, m 178°, 178-181°, 181-182°, $[\alpha]_{\text{D}}^{20}$ -83° (c 1, H₂O). Purified by recrystn from EtOH. [Horikoshi *J Biochem (Tokyo)* 35 39 1042;

Goebel and Avery *J Exptl Medicine* **50** 521 1929; Snyder and Link *J Am Chem Soc* **75** 1758.] It is a chromogenic substrate for β -galactosidases [Buoncore et al. *J Appl Biochem* **2** 390 1980].

4-Nitrophenyl- α -D-glucopyranoside [3767-28-0] **M 301.3, m 206-212°, 216-217° (sinters at 210°), $[\alpha]_D^{20} +215°$ (c 1, H₂O).** Purified by recrystn from H₂O, MeOH or EtOH. [Jermyn *Aust J Chem* **7** 202 1954; Montgomery et al. *J Am Chem Soc* **64** 690 1942.] It is a chromogenic substrate from α -glucosidases [Oliviera et al. *Anal Biochem* **113** 188 1981], and is a substrate for glucansucrases [Binder and Robyt *Carbohydr Res* **124** 287 1983]. It is a chromogenic substrate for β -glucosidases [Weber and Fink *J Biol Chem* **255** 9030 1980].

4-Nitrophenyl- β -D-glucopyranoside [2492-87-7] **M 301.2, m 164°, 164-165°, 165°, $[\alpha]_D^{20} -107°$ (c 1, H₂O).** Purified by recrystn from EtOH or H₂O. [Montgomery et al. *J Am Chem Soc* **64** 690 1942; Snyder and Link *J Am Chem Soc* **75** 1758 1953.]

Nonactin [6833-84-7] **M 737.0, m 147-148°, $[\alpha]_D^{20} 0° (\pm 2°)$ (c 1.2, CHCl₃).** This macrotetrolide antibiotic was rerystd from MeOH as colourless needles, and dries at 90°/20h/high vacuum. [*Helv Chim Acta* **38** 1445 1955, **55** 1371 1972; *Tetrahedron Lett* 3391 1975.]

N-Nonanoyl-n-methylglucamine (Mega-9) [85261-19-4] **M 335.4, m 87-89°.** A non-ionic detergent purified as *n*-decanoyl-*N*-methylglucamine above. [Hildreth *Biochem J* **207** 363 1982.]

Nonyl- β -D-glucopyranoside [69984-73-2] **M 306.4, m 67.5-70°, $[\alpha]_D^{20} -34.4°$ (c 5, H₂O), $[\alpha]_D^{25} -28.8°$ (c 1, MeOH).** Purified by recrystn from Me₂CO and stored in well stoppered containers as it is *hygroscopic*. [Pigman and Richtmyer *J Am Chem Soc* **64** 369 1942.] It is a UV transparent non-ionic detergent for solubilising membrane proteins [Schwendener et al. *Biochem Biophys Res Commun* **100** 1055 1981.]

L-Noradrenaline (Adrenor, R-2-amino-1-[3,4-dihydroxyphenyl]ethan-1-ol, L-norepinephrine) [51-41-2, 69815-49-2 (*bitartrate salt*)] **M 169.2, m 216.5-218°(dec), ~220-230°(dec), $[\alpha]_D^{20} -45°$ (c 5, N HCl), $[\alpha]_D^{25} 37.3°$ (c 5, 1 equiv aqueous HCl), $pK_1^{25} 5.58$ (phenolic OH), $pK_2^{25} 8.90$ (phenolic OH), $pK_3^{25} 9.78$ (NH₂).** Recrystd from EtOH and stored in the dark under N₂. [pKa, Lewis *Brit J Pharmacol Chemother* **9** 488 1954; UV: Bergström et al. *Acta Physiol Scand* **20** 101 1950; Fluorescence: Bowman et al. *Science NY* **122** 32 1955; Tullar *J Am Chem Soc* **70** 2067 1948.] The *L-tartrate salt monohydrate* has **m 102-104.5°, $[\alpha]_D^{25} -11°$ (c 1.6, H₂O),** after recrystn from H₂O or EtOH.

L-Noradrenaline hydrochloride (Arterenol) [329-56-6] **M 205.6, m 145.2-146.4°, ~150°(dec), $[\alpha]_D^{25} -40°$ (c 6, H₂O), pK see above.** Recrystd from isoPrOH and stored in the dark as it is oxidised in the presence of light (see preceding entry). [Tullar *J Am Chem Soc* **70** 2067 1948.]

Novobiocin (7-[O³-carbamoyl-5-O⁴-dimethyl- β -L-lyso-6-desoxyhexahydropyranosyloxy]-4-hydroxy-3[4-hydroxy-3-(3-methylbut-2-enyl)-benzyl-amino]-8-methylcoumarin) [303-81-1] **M 612.6, two forms m 152-156° and m 172-174°, 174-178°, λ_{max} at 330nm (acid EtOH), 305nm (alk EtOH), $[\alpha]_D^{25} -63°$ (c 1, EtOH), pK_1 4.03 (4.2), pK_2 9.16.** Crystd from EtOH and stored in the dark. It has also been recrystd from Me₂CO-H₂O. [Hoeksema et al. *J Am Chem Soc* **77** 6710 1955; Kaczka et al. *J Am Chem Soc* **77** 9404 1955.]

The **sodium salt** [1476-53-5] **M 634.6, m 210-215°, 215-220°(dec), 222-229°, $[\alpha]_D^{25} -38°$ (c 1, H₂O)** has been recrystd from MeOH, then dried at 60°/0.5mm. [Sensi, Gallo and Chiesa, *Anal Chem* **29** 1611 1957; Kaczka et al. *J Am Chem Soc* **78** 4126 1956.]

5'-Nucleotidase (from Electric ray, *Torpedo sp*) [9027-73-0] [**EC 3.1.3.5**], **amorphous.** Purified by dissolving in Triton X-100 and deoxycholate, and by affinity chromatography on concanavalin A-Sepharose and AMP-Sepharose [Grondal and Zimmerman *Biochem J* **245** 805 1987].

Nucleotide thiophosphate analogues. The preparation and purification of [³H]ATP γ S, [³H]GTP γ S, s⁶ITP γ S (6-thioinosine), cl⁶ITP γ S (6-chloroinosine) and [³H]ATP γ S are described and the general purification

was achieved by chromatography of the nucleotide thiophosphates in the minimum volume of H₂O placed onto a DEAE-Sephadex A25 column and eluting with a linear gradient of triethylammonium bicarbonate (0.1 to 0.6M for G and I nucleotides and 0.2 to 0.5M for A nucleotides). [*Biochim Biophys Acta* **276** 155 1972.]

Nystatin dihydrate (Mycostatin, Fungicidin) [1400-61-9] **M 962.1, m dec>160° (without melting by 250°)**, $[\alpha]_D^{25} -7^\circ$ (0.1N HCl in MeOH), -10° (AcOH), $+12^\circ$ (Me₂NCHO), $+21^\circ$ (pyridine). Light yellow powder with the following solubilities at ~28°: MeOH (1.1%), ethylene glycol (0.9%), H₂O (0.4%), CCl₄ (0.12%), EtOH (0.12%), CHCl₃ (0.05%) and *C₆H₆ (0.03%). Could be pptd from MeOH soln by addition of H₂O. Aqueous suspensions of this macrolide antifungal antibiotic are stable at 100°/10min at pH 7.0 but decomposes rapidly at pH <2 and >9, and in the presence of light and O₂. [Birch et al. *Tetrahedron Lett* 1491, 1485 1964; Weiss et al. *Antibiot Chemother* **7** 374 1957.] It contains a mixture of components A₁, A₂ and A₃.

Octyl-β-D-glucopyranoside [29836-26-8] **M 292.4, m 62-65°, 63.8-65°**, $[\alpha]_D^{20} -34^\circ$ (c 4, H₂O). Purified by recrystn from Me₂CO. It is *hygroscopic* and should be stored in a well stoppered container. [Noller and Rockwell *J Am Chem Soc* **60** 2076 1938; Pigman and Richtmyer *J Am Chem Soc* **64** 369 1942.] It is a UV transparent non-ionic dialysable detergent for solubilising membrane proteins. The α-D-isomer with $[\alpha]_D^{20} +118^\circ$ (c 1, MeOH) has similar solubilising properties. [Lazo and Quinn *Anal Biochem* **102** 68 1980; Stubbs et al. *Biochim Biophys Acta* **426** 46 1976.]

Orcine monohydrate (3,5-dihydroxytoluene) [6153-39-5] **M 142.2, m 56°, 56-58°, 58°, b 147°/5 mm, pK₁²⁰ 9.48 (9.26), pK₂²⁰ 11.20 (11.66)**. Purified by recrystn from H₂O as the monohydrate. It sublimes *in vacuo* and the *anhydrous* compound has **m 106.5-108° (110°, 108°)**. Also can be recrystd from CHCl₃ (plates) or *C₆H₆ (needles or prisms). [UV: Kiss et al. *Bull Soc Chim Fr* 275 1949; Adams et al. *J Am Chem Soc* **62** 732 1940.]

Orosomucoid (glycoprotein α₁ acid, from human plasma) [66455-27-4] **M_r 42000-44000, amorphous**. Purified by passage through a carboxymethyl cellulose column and through a Sephadex G-25 column. [Aronson et al. *J Biol Chem* **243** 4564 1968.]

Orotic acid Li salt H₂O (1-carboxy-4,6-dihydroxypyrimidine Li salt H₂O) [5266-20-6] **M 180.0, m >300°, pK₁ 2.8 (CO₂H), pK₂ 9.4 (OH), pK₃ >13 (OH) (for free acid)**. It is soluble in H₂O at 17° and 100°. Best to acidify an aqueous soln, isolating the free acid which is recrystd from H₂O (as monohydrate) **m 345-347° (345-346°)**, then dissolving in EtOH, adding an equivalent amount of LiOH in EtOH and evaporating. Its solubility in H₂O is 1.28% (17°) and 2.34% (100°). [Bachstsz *Chem Ber* **63** 1000 1930; Johnson and Shroeder *J Am Chem Soc* **54** 2941 1932; UV: Shugar and Fox *Biochim Biophys Acta* **9** 199 1952.]

Oxacillin sodium salt (5-methyl-3-phenyl-4-isoxazolympenicillin sodium salt) [1173-88-2] **M 423.4, m 188°(dec)**, $[\alpha]_D^{20} +29^\circ$ (c 1, H₂O), **pK_{Est} ~ 2.7**. This antibiotic which is stable to penicillinase is purified by recrystn from isoPrOH and dried *in vacuo*. Its solubility in H₂O at 25° is 5%. [Doyle et al. *Nature* **192** 1183 1961.]

Oxolinic acid (5-ethyl-5,8-dihydro-8-oxo-1,3-dioxolo[4,5-g]quinoline-3-carboxylic acid) [14698-29-4] **M 261.2, m 313-314°(dec), 314-316°(dec), pK_{Est} ~ 2.3**. Purified by recrystn from aqueous Me₂CO or 95% EtOH. It has UV λ_{max} 220, (255.5sh), 259.5, 268, (298sh, 311sh), 321 and 326nm [ϵ 14.8, (36.8sh), 38.4, 38.4, (6.4sh, 9.2sh), 10.8 and 11.2 x 10³]. [Kaminsky and Metzger *J Med Chem* **11** 160 1968.]

Oxytocin [50-56-6] **M 1007.2, m dec on heating, $[\alpha]_D^{22} -26.2^\circ$ (c 0.53, N AcOH)**. A cyclic nonapeptide which was purified by countercurrent distribution between solvent and buffer. It is soluble in H₂O, *n*-BuOH and isoBuOH. [Bodanszky and du Vigneaud *J Am Chem Soc* **81** 2504 1959; Cash et al. *J Med Pharm Chem* **5** 413 1962; Sakakibara et al. *Bull Chem Soc Jpn* **38** 120 1965; solid phase synthesis: Bayer and

Hagenmyer *Tetrahedron Lett* 2037 1968.] It was also synthesised on a solid phase matrix and finally purified as follows: A Sephadex G-25 column was equilibrated with the aqueous phase of a mixture of 3.5% AcOH (containing 1.5% of pyridine) + *n*-BuOH + ¹⁴C₆H₆ (2:1:1) and then the organic phase of this mixture was run through. A soln of oxytocin (100mg) in H₂O (2mL) was applied to the column which was then eluted with the organic layer of the above mixture. The fractions containing the major peak [as determined by the Folin-Lowry protein assay [Fryer et al. *Anal Biochem* 153 262 1986] were pooled, diluted with twice their vol of H₂O, evaporated to a small vol and lyophilised to give oxytocin as a pure white powder (20mg, 508 U/mg). [Ives *Can J Chem* 46 2318 1968.]

Palmitoyl coenzyme A [1763-10-6] **M 1005.9**. Possible impurities are palmitic acid, S-palmitoyl thioglycolic acid and S-palmitoyl glutathione. These are removed by placing *ca* 200mg in a centrifuge tube and extracting with Me₂CO (20mL), followed by two successive extractions with Et₂O (15mL) to remove S-palmitoyl thioglycolic acid and palmitic acid. The residue is dissolved in H₂O (4 x 4 mL), adjusted to pH 5 and centrifuged to remove insoluble S-palmitoyl glutathione and other insoluble impurities. To the clear supernatant is added 5% HClO₄ (6mL) whereby S-palmitoyl CoA pptes. The ppte is washed with 0.8% HClO₄ (10mL) and finally with Me₂CO (3 x 5mL) and dried *in vacuo*. It is stable for at least one year in dry form at 0° in a desiccator (dark). Solns are stable for several months at -15°. Its solubility in H₂O is 4%. The adenine content is used as the basis of purity with λ_{\max} at 260 and 232nm (ϵ 6.4 x 10⁶ and 9.4 x 10⁶ cm²/mol respectively). Higher absorption at 232nm would indicate other thio ester impurities, e.g. S-palmitoyl glutathione, which absorb highly at this wavelength. Also PO₄ content should be determined and acid phosphate can be titrated potentiometrically. [Seubert *Biochem Prep* 7 80 1960; Srer et al. *Biochim Biophys Acta* 33 31 1959; Kornberg and Pricer *J Biol Chem* 204 329, 345 1953.]

3-Palmitoyl-sn-glycerol (*R*-glycerol-1-palmitate, *L*- β -palmitin) [32899-41-5] **M 330.5, d^{27.3} 0.9014, m 66.5° (α -form), 74° (β' -form) and 77° (β -form)**. The stable β -form is obtained by crystn from EtOH or Skellysolve B and recrystn from Et₂O provides the β' -form. The α -form is obtained on cooling the melt. [Malkin and el Sharbagy *J Chem Soc* 1631 1936; Chapman *J Chem Soc* 58 1956; Luton and Jackson *J Am Chem Soc* 70 2446 1948.]

Pancuronium bromide (2 β ,16 β -dipiperidino-5 α -androstan-3 α ,17 β -diol diacetate dimethobromide) [15500-66-0] **M 732.7, m 212-215°, 215°**. Odourless crystals with a bitter taste which are purified through acid-washed Al₂O₃ and eluted with isoPrOH-EtOAc (3:1) to remove impurities (e.g. the monomethobromide) and eluted with isoPrOH to give the pure bromide which can be recrystd from CH₂Cl₂-Me₂CO or isoPrOH-Me₂CO. It is soluble in H₂O (50%) and CHCl₃ (3.3%) at 20°. It is a non-depolarising muscle relaxant. [Buckett et al. *J Med Chem* 16 1116 1973.]

D-Panthenol (Provitamin B, *R*-2,4-dihydroxy-3,3-dimethylbutyric acid 3-hydroxypropylamide) [81-13-0] **M 205.3, b 118-120°/0.02mm, d₂₀²⁰ 1.2, n_D²⁰ 1.4935, [α]_D²⁰ (c 5, H₂O)**. Purified by distn *in vacuo*. It is a slightly *hygroscopic* viscous oil. Soluble in H₂O and organic solvent. It is hydrolysed by alkali and strong acid. [Rabin *J Am Pharm Assoc (Sci Ed)* 37 502 1948; Bonati and Pitré *Farmaco Ed Scient* 14 43 1959.]

***R*-(+)-Pantothenic acid sodium salt** (*N*-[2,4-dihydroxy-3,3-dimethylbutyryl] β -alanine Na salt) [867-81-2] **M 241.2, [α]_D²⁵ +27.1° (c 2, H₂O), pK²⁵ 4.4 (for free acid)**. Crystd from EtOH, very *hygroscopic* (kept in sealed ampoules). The free acid is a viscous *hygroscopic* oil with [α]_D²⁵ +37.5° (c 5, H₂O), easily destroyed by acids and bases.

***R*-(+)-Pantothenic acid Ca salt** [(*D*)- 137-08-6; 63409-48-3] **M 476.5, m 195-196°, 200-201°, [α]_D²⁰ +28.2° (c 5, H₂O)**. Crysts in needles from MeOH, EtOH or isoPrOH (with 0.5mol of isoPrOH). Moderately *hygroscopic*. The *S*-benzylisothiuronium salt has **m 151-152° (149° when crystd from Me₂CO)**. [Kagan et al. *J Am Chem Soc* 79 3545 1957; Wilson et al. *J Am Chem Soc* 76 5177 1954; Stiller and Wiley *J Am Chem Soc* 63 1239 1941.]